

Effect of Dietary Supplementation of Arginine, Tryptophan, and Taurine on Productive Performance, Egg Quality, and Health Status of Laying Hens Raised Under Heat Stress Conditions

C. H. Kwon, J. H. Nam, G. P. Han, D. Y. Kim, & D. Y. Kil* Department of Animal Science and Technology, Chung-Ang University, Anseong-si, Gyeonggi-do 17546, Republic of Korea *Corresponding author: dongyong@cau.ac.kr (Received 06-04-2023; Revised 26-05-2023; Accepted 07-06-2023)

ABSTRACT

The objective of this experiment was to investigate the effect of dietary supplementation of arginine (Arg), tryptophan (Trp), and taurine (Tau) on productive performance, egg quality, liver visual characteristics, antioxidant status, immune response, and stress indicator of laying hens raised under heat stress conditions. A total of two hundred eighty 47-wk-old Hy-Line Brown laying hens were randomly allotted to 1 of 4 dietary treatments with 7 replicates consisting of 10 cages per replicate. A basal diet (BD) was prepared to meet or exceed nutrient requirement estimates. Two additional diets were formulated to increase either digestible Arg or Trp by 50% greater than the BD. Finally, one more diet was prepared by adding 0.5% Tau to the BD. The experimental diets were fed to hens on an ad libitum basis for 8 wk. Average room temperature and relative humidity were maintained at 30.7±1.41°C and 72.5±11.61%, respectively. Results indicated that laying hens in Arg and Trp treatments tended (p= 0.06) to have a higher egg yolk color (Roche color fan) than those in the Tau treatment. Likewise, there was a tendency (p= 0.05) for a lower liver color score in the Tau treatment than Arg and Trp treatments. In conclusion, dietary supplementation of Arg, Trp, and Tau at the current levels (0.37% SID Arg, 0.075% SID Trp, and 0.5% Tau) in diets has no positive effects on productive performance, egg quality, liver visual characteristics, antioxidant status, immune response, and stress indicators of laying hens raised under the current heat stress conditions.

Keywords: arginine; heat stress; laying hen; taurine; tryptophan

INTRODUCTION

Heat stress is one of the major environmental stressors in the poultry industry because of the current global warming situation (Habibian *et al.*, 2015). In particular, poultry is considered highly sensitive to heat stress conditions because of a lack of sweat glands and high feather coverage (Brugaletta *et al.*, 2022). Poultry exposed to heat stress, therefore, is well-known to show various physiological malfunctions, which leads to an impairment in productive performance, product quality, and health status (Lara & Rostagno, 2013; Sugiharto, 2020). Thus, the development of effective strategies to decrease the negative outcomes from heat stress is crucial for the current and future poultry industry.

Dietary managements by supplementation of functional nutrients have been widely practiced to reduce heat stress in the current poultry industry. Various amino acids (AA), including arginine (Arg), tryptophan (Trp), and taurine (Tau) have gained increasing attention as functional nutrients to mitigate the heat stress of poultry. Dietary Arg is an essential amino acid (EAA) for poultry, and it can act as a precursor molecule of polyamines, creatine, and nitric oxide, which are wellknown to have a variety of functions, especially for

improving productive performance, product quality, and health status in poultry (Khajali & Wideman, 2010). Similarly, dietary Trp is also an EAA for poultry. In particular, Trp is a precursor of serotonin, which is a potential stress-relieving hormonal molecule in animals exposed to stressful environments (Woodger et al., 1979; Martin et al., 2000; Shen et al., 2012). Likewise, dietary Tau is considered one of the semi-EAA for animals, reporting to show stress-alleviating effects, possibly due to its role in osmoregulation, anti-inflammation, cell membrane stabilization, antioxidation, and neuromodulation in animals (Cassol et al., 2010). Accordingly, several previous experiments reported the beneficial effects of additional supplementation of Arg, Trp, and Tau in the diets of broiler chickens (Chamruspollert et al., 2004; Yue et al., 2017; Hafeez et al., 2021). However, there is still limited information regarding the effect of this functional AA in laying hens exposed to heat stress. Moreover, comparative effects of these 3 functional AA have not been reported previously in laying hens exposed to heat stress.

Therefore, the objective of the present study was to compare the efficacy of dietary supplementation of Arg, Trp, and Tau on productive performance, egg quality, liver visual characteristics, antioxidant status, immune response, and stress indicator in laying hens raised under heat stress conditions.

MATERIALS AND METHODS

Animals, Diets, and Experimental Design

All experimental procedures were reviewed and approved by the Institutional Animal Care and Use Committee at Chung-Ang University (IACUC No. 2020-00099). A total of two hundred eighty 47-wk-old Hy-Line Brown laying hens were allotted to 1 of 4 dietary treatments with 7 replicates in a completely randomized design. Each replicate consisted of 10 consecutive cages (24cm × 36cm × 39cm) with 1 hen per cage. Initial body weight (BW) and egg production rate of hens were similar among treatments. A basal diet (BD) was prepared to meet or exceed nutrient requirement estimates for Hy-Line Brown laying hens (Table 1; Hy-Line Brown International, 2018). The concentrations of AA in the diet were based on standardized ileal digestible (SID) AA. The concentrations of Arg and Trp in the BD were 0.74% SID Arg and 0.15% SID Trp, respectively. Two additional diets were formulated to increase either SID Arg or Trp by approximately 50% greater than the BD with supplementation of 0.37% Arg or 0.075% Trp. Thus, one diet (T-Arg) contained 1.11% SID Arg, whereas the other diet (T-Trp) contained 0.225% SID Trp. Finally, one more diet (T-Tau) was prepared by adding 0.5% Tau to the BD. The continuous heat stress conditions were maintained throughout the experiment. The averages of ambient temperature and relative humidity were 30.7±1.41°C and 72.5±11.61%, respectively. Based on the heat stress index calculated from the ambient temperature and relative humidity (Hy-Line Brown International, 2016), the heat stress index in our environmental conditions was 83, indicating that laying hens in the present experiment were raised under severe heat stress conditions (Hy-Line Brown International, 2016). The experimental diets were fed to hens on an ad libitum basis for 8 wk. A 16-h lighting schedule (16 light:8 dark) was used during the entire experiment.

Sample Collection and Analysis

At the conclusion of the experiment, 3 birds with a BW that was close to the average BW in each replicate were selected. One bird was euthanized by CO_2 asphyxiation, and then immediately dissected. One of the remaining 2 birds was used for analyzing the blood heterophil to lymphocyte ratio (H:L ratio) as a stress indicator, whereas the other bird was used for analyzing immune responses.

Productive Performance

Productive performance, including hen-day egg production, egg weight (EW), egg mass, and broken and shell-less egg production rate were recorded daily. However, feed intake (FI) and feed conversion ratio (FCR) were recorded weekly. Egg mass was calculated by multiplying EW by hen-day egg production.

Egg Quality

Egg quality was assessed using randomly collected samples of 10 eggs per replicate with 5 eggs per day during the last 2 days of 4 and 8 wk. The detailed procedures for egg quality were demonstrated in our previous study (Kim *et al.*, 2017).

Table 1. Composition and nutrient contents of basal diets (asfed basis)

Items	Value %						
Ingredients							
Corn	61.50						
Soybean meal, 45% Crude protein	16.90						
Corn gluten meal	1.64						
DDGS ¹	5.00						
Tallow	0.79						
Monodicalcium phosphate	1.26						
Limestone	10.26						
54% Lysine H ₂ SO ₄	0.34						
98.5% Threonine	0.06						
Liquid Methionine	0.28						
98.5% L-Tryptophan	0.11						
NaCl	0.27						
50% Choline	0.10						
NaHCO ₃	0.10						
Vitamin premix ²	0.09						
Mineral premix ³	0.10						
Celite	1.20						
Total	100.00						
Nutrient and energy content ⁴							
AME ⁵ kcal/kg	2,740						
Crude protein %	14.61						
Digestible lysine %	0.71						
Digestible methionine + cysteine %	0.65						
Digestible methionine %	0.44						
Digestible threonine %	0.50						
Digestible tryptophan %	0.15						
Digestible arginine %	0.74						
Digestible isoleucine %	0.51						
Digestible valine %	0.65						
Total calcium %	4.18						
Available phosphorus %	0.36						

Note: ¹DDGS, corn distillers dried grains with solubles.

²Provided per kg of the complete diet: vitamin A 11,700 IU (retinyl acetate); vitamin D3 3,600 IU; vitamin E 27 IU (DL- α -tocopheryl acetate); vitamin K3 2.7 mg (menadione dimethylpyrimidinol); vitamin B1 2.7 mg; vitamin B2 6.3 mg; vitamin B6 4.5 mg; vitamin B12 18 µg; folic acid 1.35 mg; biotin 135 µg; niacin 45 mg; panto-thenic acid 10.8 mg.

 3Provided per kg of the complete diet: copper 7.35 mg; iron 46.75 mg; manganese 87.34 mg; zinc 75.21 mg; chromium 100 µg; selenium 235 µg.

⁴Calculated values from Hy-Line (2018).

⁵AMEn, nitrogen-corrected apparent metabolizable energy.

Liver Visual Characteristics

For a measure of the occurrence of fatty liver, the liver attached to the body was pictured to measure the subjective fatty liver score using a scale from 1 to 5 (1= dark red; 5= yellowish red; Choi *et al.*, 2012). In addition, the objective CIE color scale for the lightness (L*), redness (a*), and yellowness (b*) was also determined using a colorimeter (model CR-10, Konica Minolta Optics Inc., Tokyo, Japan). The liver hemorrhage was scored from 0 to 5, 0 indicating normal liver and 5 indicating large and massive hemorrhages (Diaz *et al.*, 1999).

Antioxidant Status

Antioxidant statuses in the liver, such as malondialdehyde (MDA), total antioxidant capacity (TAC), and reactive oxygen species (ROS), were determined using a commercially available OxiSelectTM TBARS Assay Kit (MDA Quantitation; STA-330, Cell Biolabs, USA), OxiSelectTM Total Antioxidant Capacity (TAC) Assay Kit (STA-360, Cell Biolabs, USA), and OxiSelectTM In Vitro ROS/RNS Assay Kit (STA-347, Cell Biolabs, USA), respectively, according to the manufacturer's protocol. Protein concentrations were also analyzed using a commercial kit (Thermo Fisher Scientific Inc.; Shen *et al.*, 2012). The detailed procedure was reported in our previous experiment (Yu *et al.*, 2021).

Immune Responses

Cutaneous basophil hypersensitivity (CBH), a measure of cell-mediated immune responses, was determined based on the method of Kean & Lamont (1994). The detailed procedure was reported in our previous experiment (Kim *et al.*, 2021).

Stress Indicators

The H:L ratio was measured as a stress biomarker. The H:L ratio in the blood was analyzed by the method of Lentfer *et al.* (2015) with a minor modification. The detailed procedure was reported in our previous experiment (Yu *et al.*, 2021).

Statistical Analysis

All data were analyzed by one-way ANOVA as a completely randomized design using the PROC MIXED procedure in SAS (SAS Institute Inc., Cary, NC, USA). The replicate was considered as an experimental unit. Data were checked for outliers using the UNIVARIATE procedure of SAS, but no outliers were identified. The LSMEANS procedure was used to calculate treatment means. Means were separated by the PDIFF option in SAS. A probability of p<0.05 was considered significant and $0.05 \le p \le 0.10$ was considered a tendency.

RESULTS

Productive Performance

During the overall experimental period, the productive performance, including final BW, BWG, hen-day egg production, EW, egg mass, FI, FCR, and broken and shell-less egg production in laying hens, was not influenced by dietary treatments (Table 2).

Egg Quality

The egg quality of laying hens was not influenced by dietary treatments (Table 3). However, laying hens fed diets supplemented with additional Arg and Trp tended (p= 0.06) to have a higher egg yolk color (Roche color fan) than those fed diets supplemented with Tau.

Liver Visual Characteristics and Antioxidant Status

The liver color and hemorrhagic score of laying hens were not influenced by dietary treatments (Table 4). However, laying hens fed diets supplemented with Tau tended (p= 0.05) to show the least liver color score among dietary treatments. Similarly, the antioxidant

Table 2. Effects of dietary supplementation of Arg, Trp, and Tau on productive performance of laying hens raised under heat stress conditions¹

Variables ³		Dietary t				
	CON	T-Arg	T-Trp	T-Tau	SEM	p-value
Initial BW, g	1,817	1,820	1,818	1,819	47.9	1.00
Final BW, g	1,806	1,832	1,787	1,814	39.6	0.88
BWG, g	-11	12	-31	-5	16.6	0.35
Hen-day egg production, %	89.4	90.3	88.7	91.8	1.51	0.31
EW, g	56.3	56.7	56.8	55.9	0.44	0.48
Egg mass, g	50.3	51.2	50.3	51.5	0.88	0.71
FI, g/hen/d	99	99	97	99	1.6	0.57
FCR, g/g	1.96	1.94	1.92	1.93	0.023	0.60
Broken and shell-less eggs, %	0.15	0.16	0.14	0.13	0.077	0.60

Note: ¹Data are least squares means of 7 replicates per treatment.

²CON: control (0.74% SID arginine + 0.15% SID tryptophan); T-Arg: arginine treatment (CON + 0.37% SID arginine); T-Trp: tryptophan treatment (CON + 0.075% SID tryptophan); T-Tau: taurine treatment (CON + 0.5% taurine).

³BW: body weight; BWG: body weight gain; EW: egg weight; FCR: feed conversion ratio; FI: feed intake.

status in the liver of laying hens was not influenced by dietary treatments (Table 5).

Immune Responses and Stress Indicators

No significant effect of dietary treatments on the CBH response was found (Table 6). Likewise, H:L ratio of hens was not affected by dietary treatments (Table 6).

DISCUSSION

Productive Performance

Heat stress is reported to decrease egg production, EW, and FI of laying hens (Barrett *et al.*, 2019; Xing *et*

al., 2019). This decrease in the productive performance of laying hens was most likely due to the decrease in FI, which is directly involved in decreasing the supply of available energy and nutrients used for the productive performance of laying hens (Mashaly *et al.*, 2004). In line with this observation, Zhou *et al.* (1998) and Mahmoud *et al.* (1996) reported that heat stress decreased plasma Ca and protein concentration, which are essential for egg formation in laying hens. Similar results for decreased nutrient and energy utilization with decreased FI were observed in broiler chickens exposed to heat stress (Bonnet *et al.*, 1997).

As a practical approach to reducing heat stress, nutritional manipulation is widely practiced in the current poultry production system (Khan *et al.*, 2012).

Table 3. Effects of dietary supplementation of Arg, Trp, and Tau on egg quality of laying hens raised under heat stress conditions¹

			Dietary treatments ²				1
variables		CON	T-Arg	T-Trp	T-Tau	SEIM	p-value
Eggshell thickness, μm		396	390	399	398	4.6	0.50
Eggshell strength, kg/cm ²		3.84	3.81	3.90	4.03	0.129	0.64
Haugh unit		88.4	88.0	88.8	87.7	0.79	0.74
Egg yolk color (Roche color fan)		7.0	7.1	7.1	6.9	0.06	0.06
Eggshell color (Shell color fan)		12.7	12.7	12.8	12.4	0.25	0.67
Eggshell color (CIE Lab value) ³	L*	53.7	53.5	53.6	54.1	0.68	0.83
	a*	20.3	20.4	20.4	20.5	0.32	0.98
	b*	30.5	30.2	30.1	30.4	0.20	0.62

Note: ¹Data are least squares means of 7 replicates per treatment.

²CON: control (0.74% SID arginine + 0.15% SID tryptophan); T-Arg: arginine treatment (CON + 0.37% SID arginine); T-Trp: tryptophan treatment (CON + 0.075% SID tryptophan); T-Tau: taurine treatment (CON + 0.5% taurine).

³L*: lightness; a*: redness; b*: yellowness.

Table 4. Effects of dietary supplementation of Arg, Trp, and Tau on liver visual characteristics of laying hens raised under heat stress conditions¹

Variables		Dietary treatments ²				CEM	
variables		CON	T-Arg	T-Trp	T-Tau	SEIVI	p-value
Liver color (CIE Lab value) ³	L*	31.7	30.4	30.9	28.8	1.17	0.35
	a*	19.8	19.2	19.9	18.3	0.77	0.47
	b*	10.6	10.3	10.3	9.2	0.93	0.71
Liver color score		2.96	2.64	2.68	1.82	0.286	0.05
Liver hemorrhagic score		0.82	1.11	1.00	0.82	0.289	0.87

Note: ¹Data are least squares means of 7 replicates per treatment.

²CON: control (0.74% SID arginine + 0.15% SID tryptophan); T-Arg: arginine treatment (CON + 0.37% SID arginine); T-Trp: tryptophan treatment (CON + 0.075% SID tryptophan); T-Tau: taurine treatment (CON + 0.5% taurine).

³L*: lightness; a*: redness; b*: yellowness.

Table 5. Effects of dietary supplementation of Arg, Trp, and Tau on antioxidant capacity of laying hens raised under heat stress conditions¹

V ₂		Dietary tr	CEM			
Variables	CON	T-Arg	T-Trp	T-Tau	SEM	p-value
MDA, µmol/mg protein	2.94	2.60	2.93	3.00	0.379	0.87
TAC, μmol/mg protein	728	751	769	738	30.8	0.81
ROS, mM	65.8	53.7	54.6	57.7	7.28	0.64

Note: ¹Data are least squares means of 7 replicates per treatment.

²CON: control (0.74% SID arginine + 0.15% SID tryptophan); T-Arg: arginine treatment (CON + 0.37% SID arginine); T-Trp: tryptophan treatment (CON + 0.075% SID tryptophan); T-Tau: taurine treatment (CON + 0.5% taurine).

³MDA: malondialdehyde; TAC: total antioxidant capacity; ROS: reactive oxygen species.

		CBH te	est mm		
Dietary treatments ²		H:L ratio ³			
	0 h	6 h	12 h	24 h	
CON	0.00	0.41	0.29	0.23	0.32
T-Arg	0.00	0.34	0.20	0.13	0.25
T-Trp	0.00	0.32	0.21	0.09	0.30
T-Tau	0.00	0.41	0.33	0.19	0.29
SEM		0.056	0.051	0.054	0.033
p-value		0.58	0.29	0.32	0.59

Table 6. Effects of dietary supplementation of Arg, Trp, and Tau on cutaneous basophil hypersensitivity (CBH) test and stress indicator of laying hens raised under heat stress conditions¹

Note: ¹Data are least squares means of 7 replicates per treatment.

²CON: control (0.74% SID arginine + 0.15% SID tryptophan); T-Arg: arginine treatment (CON + 0.37% SID arginine); T-Trp: tryptophan treatment (CON + 0.075% SID tryptophan); T-Tau: taurine treatment (CON + 0.5% taurine).

³H:L ratio: heterophil to lymphocyte ratio.

Dietary Trp is an essential amino acid for animals and is also known to decrease the stress response in animals (Koopmans et al., 2012). Dietary Trp is reported to ameliorate abnormal behavior and stress responses of animals by promoting serotonin synthesis (Shen et al., 2012). It is appreciated that serotonin is associated with regulating the central nervous system for inhibiting aggressive behavior and controlling stress responses in animals (Shen et al., 2012). However, we did not find any beneficial effects of T-Trp treatments (i.e., 50% higher than the requirement of SID Trp) in the current experiment. A similar result was also observed by Dong et al. (2012), who reported that dietary supplementation of 0.2 g/kg Trp and 0.4 g/kg Trp had no effect on the productive performance of laying hens raised under heat stress conditions. The reason for these results may be related to the fact that the current concentrations of Trp (0.225% SID Trp) in T-Trp treatments by 0.075% Trp supplementation in diets may not be sufficient to show the beneficial effects. In addition, based on the ideal protein concept, the balance of amino acid concentrations is important for the normal productive performance of laying hens. The concentration of other amino acids should be concomitantly increased with increasing levels of Trp from 0.15% SID Trp to 0.225% SID Trp when laying hens was raised under heat stress conditions (Dong et al., 2012), which was not applied in this experiment.

Dietary Arg is an essential amino acid for poultry, and it can be metabolized to various functional molecules such as polyamines, creatine, and nitric oxides, which are likely related to an amelioration in heat stress (Wu et al., 2010). Several previous studies have reported that dietary supplementation of Arg positively affected the productive performance of animals exposed to heat stress (Chamruspollert et al., 2004; Zhu et al., 2014; Yun et al., 2020). Similarly, dietary Tau has been reported to show several important biological functions in the animal body, and thus, it is now considered one of the semi-essential amino acids for animals (Ripps & Shen, 2012; Surai et al., 2020). In particular, dietary Tau plays an important role in stress alleviation by osmoregulation, anti-inflammation, and anti-oxidation (Cassol et al., 2010). Accordingly, many

previous studies reported that dietary supplementation of additional Tau may alleviate oxidative damage in broiler chickens exposed to heat stress. However, the results have been controversial (Shim et al., 2006; He et al., 2019; Lu et al., 2019b; Hafeez et al., 2021). In the current situation, however, limited information regarding the effects of dietary supplementation of Arg and Tau is available for laying hens exposed to heat stress. Therefore, we hypothesized that dietary supplementation of Arg and Tau might ameliorate the negative effect of heat stress conditions on laying hens. However, our results indicated that during 8 wk of the feeding trial, the productive performance of laying hens was not affected by T-Arg treatments (i.e., 50% higher than the requirement of SID Arg). Likewise, T-Tau treatments (i.e., adding 0.5% Tau to the BD) had no beneficial effect on productive performance in laying hens. The reason is unclear; however, it may be related to the increased requirements of Arg and Tau due to heat stress. Although we added additional Arg and Tau in the diet, it is unlikely that those supplemental levels fulfill their increased requirements of laying hens raised under the current heat stress conditions.

Egg Quality

Heat stress is known to decrease the egg quality of laying hens, such as eggshell thickness, eggshell strength, Haugh unit, egg yolk color, and eggshell color (Mahmoud et al., 1996; Balnave & Muheereza, 1997; Mashaly et al., 2004). The decrease in egg quality of laying hens was most likely due to a reduction in Ca intake by decreased FI (Mashaly et al., 2004; Franco-Jimenez et al., 2007). In addition, it was also reported that Ca utilization (Odom et al., 1986) and Ca uptake by duodenal epithelial cells (Mahmoud et al., 1996) were decreased by heat stress in laying hens (Mashaly et al., 2004). Mahmoud et al. (1996) reported that plasma Ca level was significantly decreased in laying hens when hens were exposed to heat stress. In addition, when hens are exposed to heat stress conditions, they show increased respiration rates to reduce body temperature through evaporative cooling (El Hadi & Sykes, 1982). The increased respiration rate of laying hens leads to

a reduction in blood partial pressure of CO_2 , HCO_3^- , and an increase in blood pH, which is often associated with respiratory alkalosis (Koelkebeck & Odom, 1994; Franco-Jimenez *et al.*, 2007). The higher blood pH reduces the amount of ionized Ca in the blood of laying hens (Odom *et al.*, 1986), which is the form of Ca utilized by the shell gland. Moreover, in laying hens, blood HCO_3^- plays an important role in the formation of the CaCO₃ required for eggshell formation (Franco-Jimenez & Beck, 2007).

A nutritional strategy has been suggested to relieve the negative effect of heat stress on egg quality, such as dietary supplementation of Trp (Dong et al., 2012). The positive effect of dietary supplementation of Trp has been reported by Dong et al. (2012), who observed that dietary supplementation of 0.2 g/kg Trp and 0.4 g/kg Trp improved eggshell strength of laying hens compared with those fed on the control diet. Dietary Trp is known to increase FI in animals (Woodger et al., 1979) and to improve feed utilization (Wu, 2009). Therefore, dietary Trp may improve the egg quality of laying hens raised under heat-stress conditions. On the contrary, to our knowledge, there have been no previous studies regarding the effect of dietary Arg and Tau on egg quality in laying hens raised under heat-stress conditions. However, dietary Arg should be sufficiently presented in poultry diets to support normal biological functions such as protein synthesis and growth (Khajali & Wideman, 2010). Likewise, dietary Tau is known to have several biological functions in laying hens, and it is considered one of the semi-essential amino acids in poultry diets raised under heat stress (Ripps & Shen, 2012; Surai et al., 2020). Therefore, we hypothesized that T-Arg, T-Trp, or T-Tau treatments might ameliorate the negative effect of heat stress conditions on egg quality in laying hens. However, our results indicated that T-Arg, T-Trp, or T-Tau treatments did not affect the egg quality of laying hens raised under heat-stress conditions. The reason is not clear; however, it may be attributed to the variations in animals and environmental conditions among the experiments because the extent of heat stress on poultry is influenced by both animal (e.g., age and genetics) and environmental factors (e.g., stocking density, ambient temperature and humidity, duration and the extent of heat stress, and rearing facility (Whitehead & Keller, 2003; Wasti et al., 2020).

Liver Visual Characteristics and Antioxidant Status

Several previous studies have reported that heat stress can increase fat deposition in poultry (Lu *et al.*, 2007; Emami *et al.*, 2021). The reason for increased fat deposition in poultry is caused by increased hepatic de novo lipogenesis (Lu *et al.*, 2019a). Different from the mammal, the adipose tissues of birds serve only as a fat storage site, and the liver is the main site of de novo lipogenesis in poultry (Emami *et al.*, 2021). Very low-density lipoprotein (VLDL) is the important lipoprotein transporting triglycerides from the liver to extrahepatic tissues in animals (Cryer, 1981). However, when the transportation of VLDL is impaired, the excessive lipids are retained in the liver, which causes hepatic steatoses such as fatty liver syndrome (FLS) and fatty liver hemorrhagic syndrome (FLHS) in poultry (Zhang et al., 2008b). The FLS and FLHS are frequently observed in laying hens and increasing economic losses in laying hens are often followed by abnormal lipid accumulation in the liver (Navarro-Villa et al., 2019). As a result, in heat stress conditions, the liver color and the hemorrhagic score of laying hens are reported to increase. We hypothesized that T-Arg, T-Trp, or T-Tau treatments might ameliorate the negative effect of heat stress conditions on the liver status in laying hens. However, our results indicated that T-Arg, T-Trp, or T-Tau treatments had no beneficial effect on the liver lipid status of laying hens raised under heat-stress conditions. This result may indicate that T-Arg, T-Trp, or T-Tau treatments have no effects on lipid synthesis and transportation in the liver of laying hens. Moreover, in this experiment, we used relatively younger laying hens, and therefore, abnormal lipid status is not frequently observed in this age of hens, such that those functional supplements are unlikely to show beneficial effects on the hepatic lipid status of laying hens exposed to heat stress.

Antioxidant status in animals can be measured by TAC, MDA, and ROS concentration in the body (Young, 2001; Zhang et al., 2008a; Rubio et al., 2016). The TAC is a measurement of assessing the antioxidant potentials of various biological samples (Rubio et al., 2016), with low TAC values representing increased oxidative stress (Young, 2001). The MDA is one of the final products of lipid peroxidation, with increasing MDA concentrations reflecting the increasing extent of lipid peroxidation (Zhang et al., 2008a). The ROS are free radicals and peroxides produced within the cells by increasing oxidative stress (Wasti et al., 2020). Many previous poultry experiments have consistently reported that heat stress increases oxidative stress in the whole body (Estévez, 2015; Surai et al., 2019). Abdel-Moneim et al. (2021) reported that poultry exposed to heat stress conditions elevated the body temperature and accelerated the metabolic rates, leading to an increase in ROS production. Accordingly, an increase in ROS concentrations over antioxidant capacity facilitates free radical-mediated oxidative chain reactions in cell components such as proteins (Stadtman & Levine, 2000), lipids (Rubbo et al., 1994), polysaccharides (Kaur & Halliwell, 1994), and deoxyribonucleic acid (LeDoux et al., 1999), which ultimately results in a decrease in cellular functions and eventually cell death.

Dietary Arg can be metabolized to polyamines, which are considered important biomolecules against oxidative stress in animals (Seiler, 1996; Wu *et al.*, 2010; Miller-Fleming *et al.*, 2015). Similarly, dietary Tau plays a role in stress alleviation by osmoregulation, antiinflammation, cell membrane stabilization, and antioxidation in animals (Cassol *et al.*, 2010). Accordingly, many previous studies reported that dietary supplementation of additional Tau may alleviate oxidative damage induced by stressors in animals (Yang *et al.*, 2015; Zhang *et al.*, 2017). Dietary Trp is known to protect tissues from oxidative damage in animals (Christen *et al.*, 1990; Reyes-Gonzales *et al.*, 2009; Del Angel-Meza *et al.*, 2011). However, our results indicated that T-Arg, T-Trp, or T-Tau treatments have no beneficial effects on the antioxidant status of laying hens raised under heat-stress conditions. The reason is unclear; however, it may be related to the age of the hens used in this experiment because antioxidant capacity, changes with age (Gu *et al.*, 2021). We used the relatively younger laying hens with high antioxidant capacity such that dietary supplementation of Trp, Arg, and Tau may show little beneficial effects on antioxidant status in laying hens exposed to heat stress.

Immune Responses and Stress Indicators

Heat stress is well-known to impair the immune system of poultry, and therefore, heat stress may increase the susceptibility of birds to pathogenic infections or diseases (Habibian *et al.*, 2014; Akhavan-Salamat & Ghasemi, 2016; Hosseini-Vashan *et al.*, 2016). Moreover, the levels of antibodies and white blood cells were reported to be reduced in heat-stressed birds (Bartlett & Smith, 2003; Mashaly *et al.*, 2004). In the current experiment, the CBH responses were measured as cell-mediated immune responses in laying hens exposed to heat stress (Kim *et al.*, 2021). The CBH responses were determined by the level of basophil infiltration at the phytohaemagglutinin P injected site of the poultry' skin (Kim *et al.*, 2021; Yu *et al.*, 2021).

Dietary Arg is a precursor of nitric oxide, which plays a role in the activity of the autonomic and central neural systems (Moncada et al., 1991; Malyshev et al., 1999), thereby affecting immune responses in animals (Moncada et al., 1991; Deroee et al., 2009). Dietary Tau is reported to improve immune functions in poultry (Lee et al., 2004; Koven et al., 2016). Similarly, dietary Trp can be used as a precursor molecule for serotonin and melatonin synthesis, which shows promoting effects on animal immune responses (Reiter, 1998; Li et al., 2011). However, limited information regarding the effects of dietary supplementation of Arg, Trp, or Tau on immune responses is available for laying hens raised at heat stress conditions. Therefore, we hypothesized that T-Arg, T-Trp, or T-Tau treatments might improve the immune response of laying hens raised under heatstress conditions. However, our results indicated that T-Arg, T-Trp, or T-Tau treatments had no beneficial effect on the immune response of laying hens raised under heat-stress conditions.

The blood H:L ratio has been widely measured as a stress biomarker in poultry (Gross & Siegel, 1983). Poultry exposed to heat stress shows an increased H:L ratio in the blood (Shini *et al.*, 2008; Akhavan-Salamat & Ghasemi, 2016). The reason for the increased H:L ratio due to heat stress is related to increasing corticosterone concentrations in the blood because circulating corticosterone affects the characteristics and function of immune cells, such as heterophils and lymphocytes (Shini *et al.*, 2008; Akhavan-Salamat & Ghasemi, 2016).

Dietary Trp is the precursor of serotonin that has many key functions in the nerve system of poultry to reduce aggressive activity, such as feather-pecking behavior, and modulate stress response (Laycock & Ball,

1990; Shea et al., 1990). Similarly, dietary Arg synthesizes nitric oxide, which regulates the production of serotonin in broiler chickens (Wideman et al., 2013). Oxidative stress is considered to be the major reason for negative outcomes for heat-stressed poultry (Akbarian et al., 2016; Farag & Alagawany, 2018). Many previous studies reported that dietary supplementation of additional Tau alleviated oxidative damages induced by stressors in broiler chickens raised under heat stress conditions (Yang et al., 2015; Zhang et al., 2017). It is suggested, moreover, that the requirement of Arg, Trp, and Tau may be increased during heat stress exposure in poultry (Surai et al., 2019). Therefore, three functional AAs were expected to exert heat stress-relieving effects on laying hens with decreasing stress indicators such as blood H:L ratio. However, in the current experiment, we found no anti-stress responses in laying hens by feeding diets supplemented with 0.37% Arg, 0.075% Trp, or 0.5% Tau. To our best knowledge, there are no data pertaining to the effects of dietary supplementation of Arg, Trp, and Tau on stress indicators such as blood H:L ratio in laying hens exposed to heat stress conditions, and therefore, it may be speculated that the current supplemental levels and its final concentrations of Arg, Trp, and Tau in diets may not be adequate to exert the beneficial effects on stress responses of laying hens as observed in other measurements.

CONCLUSION

Dietary supplementation of Arg, Trp, and Tau at the current supplemental levels (0.37% SID Arg, 0.075% SID Trp, and 0.5% Tau) has no positive effects on productive performance, egg quality, liver visual characteristics, antioxidant status, immune response, and stress indicator of laying hens raised under the current heat stress conditions.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial, personal, or other relationships with other people or organizations related to the material discussed in the manuscript.

ACKNOWLEDGEMENT

This research was carried out with the support of the Cooperative Research Program for Agriculture Science and Technology Development (Project No. PJ01502301), Rural Development Administration, Republic of Korea.

REFERENCES

Abdel-Moneim, A. M. E., A. M. Shehata, R. E. Khidr, V. K. Paswan, N. S. Ibrahim, A. A. El-Ghoul, S. A. Aldhumri, S. A. Gabr, N. M. Mesalam, A. M. Elbaz, M. A. Elsayed, M. M. Wakwak, & T. A. Ebeid. 2021. Nutritional manipulation to combat heat stress in poultry–A comprehensive review. J. Therm. Biol. 98:102915. https://doi.org/10.1016/j. jtherbio.2021.102915

- Akbarian, A., J. Michiels, J. Degroote, M. Majdeddin, A. Golian,
 & S. De Smet. 2016. Association between heat stress and oxidative stress in poultry: Mitochondrial dysfunction and dietary interventions with phytochemicals. J. Anim. Sci. Biotechnol. 7:37. https://doi.org/10.1186/s40104-016-0097-5
- Akhavan-Salamat, H. & H. A. Ghasemi. 2016. Alleviation of chronic heat stress in broilers by dietary supplementation of betaine and turmeric rhizome powder: Dynamics of performance, leukocyte profile, humoral immunity, and antioxidant status. Trop. Anim. Health Prod. 48:181-188. https://doi.org/10.1007/s11250-015-0941-1
- Balnave, D. & S. K. Muheereza. 1997. Improving eggshell quality at high temperatures with dietary sodium bicarbonate. Poult. Sci. 76:588-593. https://doi.org/10.1093/ ps/76.4.588
- Barrett, N. W., K. Rowland, C. J. Schmidt, S. J. Lamont, M. F. Rothschild, C. M. Ashwell, & M. E. Persia. 2019. Effects of acute and chronic heat stress on the performance, egg quality, body temperature, and blood gas parameters of laying hens. Poult. Sci. 98:6684-6692. https://doi. org/10.3382/ps/pez541
- Bartlett, J. R. & M. O. Smith. 2003. Effects of different levels of zinc on the performance and immunocompetence of broilers under heat stress. Poult. Sci. 82:1580-1588. https:// doi.org/10.1093/ps/82.10.1580
- Bonnet, S., P. A. Geraert, M. Lessire, B. Carré, & S. Guillaumin. 1997. Effect of high ambient temperature on feed digestibility in broilers. Poult. Sci. 76:857-863. https://doi. org/10.1093/ps/76.6.857
- Brugaletta, G., J. Teyssier, S. J. Rochell, S. Dridi, & F. Sirri. 2022. A review of heat stress in chickens. Part I: Insights into physiology and gut health. Front. Physiol. 13:934381. https://doi.org/10.3389/fphys.2022.934381
- Cassol, O. J., G. T. Rezin, F. C. Petronilho, G. Scaini, C. L. Gonçalves, G. K. Ferreira, R. Roesler, G. Schwartsmann, F. Dal-Pizzol, & E. L. Streck. 2010. Effects of N-acetylcysteine/deferoxamine, taurine and RC-3095 on respiratory chain complexes and creatine kinase activities in rat brain after sepsis. Neurochem. Res. 35:515-521. https://doi.org/10.1007/s11064-009-0089-3
- Chamruspollert, M., G. M. Pesti, & R. I. Bakalli. 2004. Influence of temperature on the arginine and methionine requirements of young broiler chicks. J. Appl. Poult. Res. 13:628-638. https://doi.org/10.1093/japr/13.4.628
- Choi, Y. I., H. J. Ahn, B. K. Lee, S. T. Oh, B. K. An, & C. W. Kang. 2012. Nutritional and hormonal induction of fatty liver syndrome and effects of dietary lipotropic factors in egg-type male chicks. Asian-Australas. J. Anim. Sci. 25:1145-1152. https://doi.org/10.5713/ajas.2011.11418
- Christen, S., E. Peterhans, & R. Stocker. 1990. Antioxidant activities of some tryptophan metabolites: possible implication for inflammatory diseases. PNAS. Nexus 87:2506-2510. https://doi.org/10.1073/pnas.87.7.2506
- Cryer, A. 1981. Tissue lipoprotein lipase activity and its action in lipoprotein metabolism. Int. J. Biochem. 13:525-541. https://doi.org/10.1016/0020-711X(81)90177-4
- Del Angel-Meza, A. R., A. J. Dávalos-Marín, L. L. Ontiveros-Martinez, G. G. Ortiz, C. Beas-Zarate, V. Chaparro-Huerta, B. M. Torres-Mendoza, & O. K. Bitzer-Quintero. 2011. Protective effects of tryptophan on neuro-inflammation in rats after administering lipopolysaccharide. Biomed. Pharmacother. 65:215-219. https://doi.org/10.1016/j. biopha.2011.02.008
- Deroee, A. F., M. Naraghi, A. F. Sontou, M. R. Ebrahimkhani, & A. R. Dehpour. 2009. Nitric oxide metabolites as biomarkers for follow-up after chronic rhinosinusitis surgery. Am. J. Rhinol. Allergy 23:159-161. https://doi. org/10.2500/ajra.2009.23.3289

Diaz, G. J., E. J. Squires, & R. J. Julian. 1999. The use of selected

plasma enzyme activities for the diagnosis of fatty liverhemorrhagic syndrome in laying hens. Avian Dis. 768-773. https://doi.org/10.2307/1592746

- Dong, X. Y., M. M. Azzam, W. Rao, D. Y. Yu, & X. T. Zou. 2012. Evaluating the impact of excess dietary tryptophan on laying performance and immune function of laying hens reared under hot and humid summer conditions. Br. Poult. Sci. 53:491-496. https://doi.org/10.1080/00071668.20 12.719149
- El Hadi, H. & A. H. Sykes. 1982. Thermal panting and respiratory alkalosis in the laying hen. Br. Poult. Sci. 23:49-57. https://doi.org/10.1080/00071688208447928
- Emami, N. K., U. Jung, B. Voy, & S. Dridi. 2021. Radical response: effects of heat stress-induced oxidative stress on lipid metabolism in the avian liver. Antioxidants 10:35. https://doi.org/10.3390/antiox10010035
- Estévez, M. 2015. Oxidative damage to poultry: from farm to fork. Poult. Sci. 94:1368-1378. https://doi.org/10.3382/ps/ pev094
- Farag, M. R. & M. Alagawany. 2018. Physiological alterations of poultry to the high environmental temperature. J. Therm. Biol. 76:101-106. https://doi.org/10.1016/j. jtherbio.2018.07.012
- Franco-Jimenez, D. J. & M. M. Beck. 2007. Physiological changes to transient exposure to heat stress observed in laying hens. Poult. Sci. 86:538-544. https://doi.org/10.1093/ ps/86.3.538
- Franco-Jimenez, D. J., S. E. Scheideler, R. J. Kittok, T. M. Brown-Brandl, L. R. Robeson, H. Taira, & M. M. Beck. 2007. Differential effects of heat stress in three strains of laying hens. J. Appl. Poult. Res. 16:628-634. https://doi. org/10.3382/japr.2005-00088
- Gross, W. B. & H. S. Siegel. 1983. Evaluation of the heterophil/ lymphocyte ratio as a measure of stress in chickens. Avian Dis. 972-979. https://doi.org/10.2307/1590198
- Gu, Y. F., Y. P. Chen, R. Jin, C. Wang, C. Wen, & Y. M. Zhou. 2021. Age-related changes in liver metabolism and antioxidant capacity of laying hens. Poult. Sci. 100:101478. https://doi.org/10.1016/j.psj.2021.101478
- Habibian, M., G. Sadeghi, S. Ghazi, & M. M. Moeini. 2015. Selenium as a feed supplement for heat-stressed poultry: A review. Biol. Trace Elem. Res. 165:183-193. https://doi. org/10.1007/s12011-015-0275-x
- Habibian, M., S. Ghazi, M. M. Moeini, & A. Abdolmohammadi. 2014. Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermoneutral or heat stress conditions. Int. J. Biometeorol. 58:741-752. https://doi.org/10.1007/ s00484-013-0654-y
- Hafeez, A., W. Akram, A. Sultan, Y. Konca, T. Ayasan, S. Naz, W. Shahzada, & R. U. Khan. 2021. Effect of dietary inclusion of taurine on performance, carcass characteristics and muscle micro-measurements in broilers under cyclic heat stress. Ital. J. Anim. Sci. 20:872-877. https://doi.org/10 .1080/1828051X.2021.1921627
- He, X., Z. Lu, B. Ma, L. Zhang, J. Li, Y. Jiang, G. Zhou, & F. Gao. 2019. Effects of dietary taurine supplementation on growth performance, jejunal morphology, appetite-related hormones, and genes expression in broilers subjected to chronic heat stress. Poult. Sci. 98:2719-2728. https://doi. org/10.3382/ps/pez054
- Hosseini-Vashan, S. J., A. Golian, & A. Yaghobfar. 2016. Growth, immune, antioxidant, and bone responses of heat stress-exposed broilers fed diets supplemented with tomato pomace. Int. J. Biometeorol. 60:1183-1192. https:// doi.org/10.1007/s00484-015-1112-9
- **Hy-Line Brown International.** 2016. Understanding Heat Stress in Layers. Hy-Line International.

- **Hy-Line Brown International.** 2018. Management Guide. Hy-Line International.
- Kaur, H. & B. Halliwell. 1994. Evidence for nitric oxidemediated oxidative damage in chronic inflammation Nitrotyrosine in serum and synovial fluid from rheumatoid patients. FEBS Lett. 350:9-12. https://doi. org/10.1016/0014-5793(94)00722-5
- Khajali, F. & R. F. Wideman. 2010. Dietary arginine: metabolic, environmental, immunological and physiological interrelationships. Worlds Poult. Sci. J. 66:751-766. https:// doi.org/10.1017/S0043933910000711
- Khan, R. U., S. Naz, Z. Nikousefat, M. Selvaggi, V. Laudadio, & V. Tufarelli. 2012. Effect of ascorbic acid in heat-stressed poultry. Worlds Poult. Sci. J. 68:477-490. https://doi. org/10.1017/S0043933910000711
- Kean, R. P. & S. J. Lamont. 1994. Effect of injection site cutaneous basophil hypersensitivity response to phytohemagglutinin. Poult. Sci. 73:1763-1765. https://doi. org/10.3382/ps.0731763
- Kim, D. Y., J. H. Kim, W. J. Choi, G. P. Han, & D. Y. Kil. 2021. Comparative effects of dietary functional nutrients on growth performance, meat quality, immune responses, and stress biomarkers in broiler chickens raised under heat stress conditions. Anim. Biosci. 34:1839. https://doi. org/10.5713/ab.21.0230
- Kim, J. H., F. M. Pitargue, H. Jung, G. P. Han, H. S. Choi, & D. Y. Kil. 2017. Effect of superdosing phytase on productive performance and egg quality in laying hens. Asian-Australas. J. Anim. Sci. 30:994. https://doi.org/10.5713/ ajas.17.0149
- Koelkebeck, K. W. & T. W. Odom. 1994. Laying hen responses to acute heat stress and carbon dioxide supplementation:
 I. Blood gas changes and plasma lactate accumulation. Comp. Biochem. Physiol. A Physiol. 107:603-606. https:// doi.org/10.1016/0300-9629(94)90358-1
- Koopmans, S. J., F. J. Van der Staay, N. Le Floc'H, R. Dekker, J. T. M. van Diepen, & A. J. M. Jansman. 2012. Effects of surplus dietary L-tryptophan on stress, immunology, behavior, and nitrogen retention in endotoxemic pigs. Anim. Sci. J. 90:241-251. https://doi.org/10.2527/ jas.2010-3372
- Koven, W., A. Peduel, M. Gada, O. Nixon, & M. Ucko. 2016. Taurine improves the performance of white grouper juveniles (*Epinephelus Aeneus*) fed a reduced fish meal diet. Aquaculture 460:8-14. https://doi.org/10.1016/j. aquaculture.2016.04.004
- Lara, L. J. & M. H. Rostagno. 2013. Impact of heat stress on poultry production. Animals 3:356-369. https://doi. org/10.3390/ani3020356
- Laycock, S. R. & R. O. Ball. 1990. Alleviation of hysteria in laying hens with dietary tryptophan. Can. J. Vet. Res. 54:291.
- LeDoux, S. P., W. J. Driggers, B. S. Hollensworth, & G. L. Wilson. 1999. Repair of alkylation and oxidative damage in mitochondrial DNA. Mutat. Res. DNA Repair (Amst). 434:149-159. https://doi.org/10.1016/S0921-8777(99)00026-9
- Lee, D. N., Y. H. Cheng, Y. S. Chuang, J. L. Shive, Y. M. Lian, H. W. Wei, & C. F. Weng. 2004. Effects of dietary taurine supplementation on growth performance, serum constituents and antibody production of broilers. Asian-Australas. J. Anim. Sci. 17:109-115. https://doi.org/10.5713/ ajas.2004.109
- Lentfer, T. L., H. Pendl, S. G. Gebhardt-Henrich, E. K. F. Frohlich, & E. Von Borell. 2015. H/L ratio as a measurement of stress in laying hens – methodology and reliability. Br. Poult. Sci. 56:157-163. https://doi.org/10.108 0/00071668.2015.1008993
- Li, N., J. E. Ghia, H. Wang, J. McClemens, F. Cote, Y. Suehiro, J. Mallet, & W. I. Khan. 2011. Serotonin activates dendritic

cell function in the context of gut inflammation. Am. J. Clin. Pathol. 178:662-671. https://doi.org/10.1016/j. ajpath.2010.10.028

- Lu, Q., J. Wen, & H. Zhang. 2007. Effect of chronic heat exposure on fat deposition and meat quality in two genetic types of chicken. Poult. Sci. 86:1059-1064. https://doi.org/10.1093/ ps/86.6.1059
- Lu, Z., X. F. He, B. B. Ma, L. Zhang, J. L. Li, Y. Jiang, G. H. Zhou, & F. Gao. 2019a. Increased fat synthesis and limited apolipoprotein B cause lipid accumulation in the liver of broiler chickens exposed to chronic heat stress. Poult. Sci. 98:3695-3704. https://doi.org/10.3382/ps/pez056
- Lu, Z., X. F. He, B. B. Ma, L. Zhang, J. L. Li, Y. Jiang, G. H. Zhou, & F. Gao. 2019b. The alleviative effects and related mechanisms of taurine supplementation on growth performance and carcass characteristics in broilers exposed to chronic heat stress. Poult. Sci. 98:878-886. https://doi. org/10.3382/ps/pey433
- Mahmoud, K. Z., M. M. Beck, S. E. Scheideler, M. F. Forman, K. P. Anderson, & S. D. Kachman. 1996. Acute high environmental temperature and calcium-estrogen relationships in the hen. Poult. Sci. 75:1555-1562. https:// doi.org/10.3382/ps.0751555
- Malyshev, I. Y., T. A. Zenina, L. Y. Golubeva, V. A. Saltykova, E. B. Manukhina, V. D. Mikoyan, L. N. Kubrina, & A. F. Vanin. 1999. NO-dependent mechanisms of adaptation to hypoxia. Nitric Oxide. 3:105-113. https://doi.org/10.1006/ niox.1999.0213
- Martin, C. L., M. Duclos, S. Aguerre, P. Mormede, G. Manier,
 & F. Chaouloff. 2000. Corticotropic and serotonergic responses to acute stress with/without prior exercise training in different rat strains. Acta Physiol. Scand. 168:421-430. https://doi.org/10.1046/j.1365-201x.2000.00683.x
- Mashaly, M. M., G. L. Hendricks 3rd, M. A. Kalama, A. E. Gehad, A. O. Abbas, & P. H. Patterson. 2004. Effect of heat stress on production parameters and immune responses of commercial laying hens. Poult. Sci. 83:889-894. https://doi.org/10.1093/ps/83.6.889
- Miller-Fleming, L., V. Olin-Sandoval, K. Campbell, & M. Ralser. 2015. Remaining mysteries of molecular biology: The role of polyamines in the cell. J. Mol. Biol. 427:3389-3406. https://doi.org/10.1016/j.jmb.2015.06.020
- Moncada, S., E. A. Higgs, H. F. Hodson, R. G. Knowles, P. Lopez-Jaramillo, T. McCall, R. M. J. Palmer, M. W. Radomski, D. D. Rees, & R. Schulz. 1991. The L-arginine: nitric oxide pathway. N. Engl. J. Med. 17:S1-S9. https://doi. org/10.1056/NEJM199312303292706
- Navarro-Villa, A., J. H. Mica, J. de los Mozos, L. A. den Hartog, & A. I. García-Ruiz. 2019. Nutritional dietary supplements to reduce the incidence of fatty liver syndrome in laying hens and the use of spectrophotometry to predict liver fat content. J. Appl. Poult. Res. 28:435-446. https://doi. org/10.3382/japr/pfz005
- Odom, T. W., P. C. Harrison, & W. G. Bottje. 1986. Effects of thermal-induced respiratory alkalosis on blood ionized calcium levels in the domestic hen. Poult. Sci. 65:570-573. https://doi.org/10.3382/ps.0650570
- Reiter, R. J. 1998. Oxidative damage in the central nervous system: protection by melatonin. Prog. Neurobiol. 56:359-384. https://doi.org/10.1016/S0301-0082(98)00052-5
- Reyes-Gonzales, M. C., L. Fuentes-Broto, E. Martínez-Ballarín, F. J. Miana-Mena, C. Berzosa, F. A. García-Gil, M. Aranda, & J. J. García. 2009. Effects of tryptophan and 5-hydroxytryptophan on the hepatic cell membrane rigidity due to oxidative stress. J. Membr. Biol. 231:93-99. https://doi.org/10.1007/s00232-009-9208-y
- Ripps, H. & W. Shen. 2012. Taurine: a "very essential" amino acid. Mol. Vis. 18:2673.
- Rubbo, H., R. Radi, M. Trujillo, R. Telleri, B. Kalyanaraman,

S. Barnes, M. Kirk, & B. A. Freeman. 1994. Nitric oxide regulation of superoxide and peroxynitrite-dependent lipid peroxidation. Formation of novel nitrogen-containing oxidized lipid derivatives. J. Biochem. Physiol. 269:26066-26075. https://doi.org/10.1016/S0021-9258(18)47160-8

- Rubio, C. P., J. Hernández-Ruiz, S. Martinez-Subiela, A. Tvarijonaviciute, & J. J. Ceron. 2016. Spectrophotometric assays for total antioxidant capacity (TAC) in dog serum: An update. BMC Vet. Res. 12:1-7. https://doi.org/10.1186/ s12917-016-0792-7
- Seiler, N. 1996. Roles of polyamines in cell biology. Principles Medical Biology 4:329-348. https://doi.org/10.1016/ S1569-2582(96)80100-0
- Shea, M. M., J. A. Mench, & O. P. Thomas. 1990. The effect of dietary tryptophan on aggressive behavior in developing and mature broiler breeder males. Poult. Sci. 69:1664-1669. https://doi.org/10.3382/ps.0691664
- Shen, Y. B., G. Voilqué, J. Odle, & S. W. Kim. 2012. Dietary L-tryptophan supplementation with reduced large neutral amino acids enhances feed efficiency and decreases stress hormone secretion in nursery pigs under social-mixing stress. J. Nutr. 142:1540-1546. https://doi.org/10.3945/ jn.112.163824
- Shim, K. S., K. T. Hwang, M. W. Son, & G. H. Park. 2006. Lipid metabolism and peroxidation in broiler chicks under chronic heat stress. Asian-Australas. J. Anim. Sci. 19:1206-1211. https://doi.org/10.5713/ajas.2006.1206
- Shini, S., P. Kaiser, A. Shini, & W. L. Bryden. 2008. Differential alterations in ultrastructural morphology of chicken heterophils and lymphocytes induced by corticosterone and lipopolysaccharide. Vet. Immunol. Immunopathol. 122:83-93. https://doi.org/10.1016/j.vetimm.2007.10.009
- Stadtman, E. R. & R. L. Levine. 2000. Protein oxidation. Ann N. Y. Acad. Sci. 899:191-208. https://doi. org/10.1111/j.1749-6632.2000.tb06187.x
- Sugiharto, S. 2020. Alleviation of heat stress in broiler chicken using turmeric (*Curcuma longa*) – a short review. Journal Animal Behaviour Biometeorology 8:215-222. https://doi. org/10.31893/jabb.20028
- Surai, P. F., I. I. Kochish, & M. T. Kidd. 2020. Taurine in poultry nutrition. Anim. Feed. Sci. Technol. 260:114339. https://doi. org/10.1016/j.anifeedsci.2019.114339
- Surai, P. F., I. I. Kochish, V. I. Fisinin, & M. T. Kidd. 2019. Antioxidant defence systems and oxidative stress in poultry biology: An update. Antioxidants 8:235. https:// doi.org/10.3390/antiox8070235
- Wasti, S., N. Sah, & B. Mishra. 2020. Impact of heat stress on poultry health and performances, and potential mitigation strategies. Animals 10:1266. https://doi.org/10.3390/ ani10081266
- Whitehead, C. C. & T. Keller. 2007. An update on ascorbic acid in poultry. 2007. World's Poult. Sci. J. 59:161-184. https:// doi.org/10.1079/WPS20030010
- Wideman, R. F., D. D. Rhoads, G. F. Erf, & N. B. Anthony. 2013. Pulmonary arterial hypertension (ascites syndrome) in broilers: A review. Poult. Sci. 92:64-83. https://doi. org/10.3382/ps.2012-02745
- Woodger, T. L., A. N. N. A. Sirek, & G. H. Anderson. 1979. Diabetes, dietary tryptophan, and protein intake regulation in weanling rats. Am. J. Physiol. Regul. Integr.

Comp. Physiol. 236:R307-R311. https://doi.org/10.1152/ ajpregu.1979.236.5.R307

- Wu, G. 2009. Amino acids: metabolism, functions, and nutrition. Amino Acids 37:1-17. https://doi.org/10.1007/ s00726-009-0269-0
- Wu, X., Z. Ruan, Y. Gao, Y. Yin, X. Zhou, L. Wang, M. Geng, Y. Hou, & G. Wu. 2010. Dietary supplementation with L-arginine or N-carbamylglutamate enhances intestinal growth and heat shock protein-70 expression in weanling pigs fed a corn-and soybean meal-based diet. Amino Acids 39:831-839. https://doi.org/10.1007/s00726-010-0538-y
- Xing, S., X. Wang, H. Diao, M. Zhang, Y. Zhou, & J. Feng. 2019. Changes in the cecal microbiota of laying hens during heat stress in mainly associated with reduced feed intake. Poult. Sci. 98:5257-5264. https://doi.org/10.3382/ps/pez440
- Yang, J., X. Zong, G. Wu, S. Lin, Y. Feng, & J. Hu. 2015. Taurine increases testicular function in aged rats by inhibiting oxidative stress and apoptosis. Amino Acids 47:1549-1558. https://doi.org/10.1007/s00726-015-1995-0
- Young, I. S. 2001. Measurement of total antioxidant capacity. J. Clin. Pathol. 54. https://doi.org/10.1136/jcp.54.5.339
- Yu, D. G., N. Namgung, J. H. Kim, S. Y. Won, W. J. Choi, & D. Y. Kil. 2021. Effects of stocking density and dietary vitamin C on performance, meat quality, intestinal permeability, and stress indicators in broiler chickens. J. Anim. Sci. Technol. 63:815. https://doi.org/10.5187/jast.2021.e77
- Yue, Y., Y. Guo, & Y. Yang. 2017. Effects of dietary L-tryptophan supplementation on intestinal response to chronic unpredictable stress in broilers. Amino Acids. 49:1227-1236. https://doi.org/10.1007/s00726-017-2424-3
- Yun, W., M. H. Song, J. H. Lee, H. J. Oh, J. S. An, G. M. Kim, S. D. Lee, S. H. Lee, H. B. Kim, & J. H. Cho. 2020. Arginine addition in a diet for weaning pigs can improve the growth performance under heat stress. J. Anim. Sci. Technol. 62:460. https://doi.org/10.5187/jast.2020.62.4.460
- Zhang, H. J., Y. M. Guo, Y. D. Tian, & J. M. Yuan. 2008a. Dietary conjugated linoleic acid improves antioxidant capacity in broiler chicks. Br. Poult. Sci. 49:213-221. https://doi. org/10.1080/00071660801989836
- Zhang, J., D. Chen, & B. Yu. 2008b. Effect of different dietary energy sources on induction of fatty liver-hemorrhagic syndrome in laying hens. Int. J. Poult. Sci. 7:1232-1236. https://doi.org/10.3923/ijps.2008.1232.1236
- Zhang, Z., L. Zhao, Y. Zhou, X. Lu, Z. Wang, J. Wang, & W. Li. 2017. Taurine ameliorated homocysteine-induced H9C2 cardiomyocyte apoptosis by modulating endoplasmic reticulum stress. Apoptosis 22:647. https://doi.org/10.1007/ s10495-017-1351-9
- Zhou, W. T., M. Fujita, S. Yamamoto, K. Iwasaki, R. Ikawa, H. Oyama, & H. Horikawa. 1998. Effects of glucose in drinking water on the changes in whole blood viscosity and plasma osmolality of broiler chickens during high temperature exposure. Poult. Sci. 77:644-647. https://doi. org/10.1093/ps/77.5.644
- Zhu, W., W. Jiang, & L. Y. Wu. 2014. Dietary L-arginine supplement alleviates hepatic heat stress and improves feed conversion ratio of Pekin ducks exposed to high environmental temperature. J. Anim. Physiol. Anim. Nutr. 98:1124-1131. https://doi.org/10.1111/jpn.12195