

PHYTOPLANKTON COMPOSITION AND ITS RELATION TO PEARL OYSTER AQUACULTURE (*Pinctada maxima*) IN SEKOTONG, WEST NUSA TENGGARA

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ABSTRACT

Pearl oyster aquaculture is one of the marine industries that is currently growing in West Nusa Tenggara. Pearl oyster (*Pinctada maxima*) is a common species of oyster that cultivated in West Nusa Tenggara waters. A suitable location is an important factor for the success of pearl aquaculture industry. It needs a good water quality and supported by appropriate physical parameters for other marine biotas. The aim of this research was to analyze phytoplankton composition (biological parameter) and its correlation to the Sekotong water condition for pearl farming production. The result showed that the pearl farming location has good water condition according to physio-chemical parameter and water nutrients. All parameters were in suitable range for the growth and development of pearl oysters.

Keywords: pearl farming, phytoplankton, *Pinctada maxima*, West Nusa Tenggara

ABSTRAK

*Budidaya Mutiara Tiram merupakan salah satu industri kelautan yang sedang berkembang di Nusa Tenggara Barat. Mutiara Tiram (*Pinctada maxima*) adalah spesies umum tiram yang dibudidayakan di perairan Nusa Tenggara Barat. Lokasi budidaya yang cocok bagi tiram merupakan faktor penting bagi keberhasilan industri budidaya mutiara. Hal ini membutuhkan kualitas air yang baik dan didukung oleh parameter fisik yang sesuai untuk biota laut lainnya. Tujuan dari penelitian ini adalah untuk menganalisis komposisi fitoplankton (parameter biologi) dan korelasinya dengan kondisi air Sekotong untuk produksi budidaya mutiara. Hasil penelitian menunjukkan bahwa lokasi budidaya mutiara memiliki kondisi air yang baik sesuai dengan parameter fisika-kimia dan nutrisi air. Semua parameter berada pada kisaran yang cocok untuk pertumbuhan dan perkembangan Mutiara Tiram.*

Kata kunci: tambak tiram, fitoplankton, *Pinctada maxima*, Nusa Tenggara Barat

I. INTRODUCTION

Indonesia has a lot of marine potential that can be developed. Pearl oyster farming is of the great potential that must be organized and utilized. *Pinctada maxima* are one of the marine organisms that found in waters of Indonesia. One of the export commodities from Indonesia is pearl that produced from *Pinctada maxima* (Anggoro, 2004).

Lombok is located in West Nusa Tenggara that develops pearl farming. With production of 600 kg per year, pearl has become iconic in Lombok. The pearl quality is known worldwide. There are 36 pearl com-

panies scattered in West Nusa Tenggara (As-tuti and Hendra, 2009).

Location plays a key role for the success of pearl farming. Good location for pearl farming must meet the standard including: water quality, nutrient, seed, infrastructure, safety and transportation (Agusta, 2012).

Pearl oyster belongs to the bivalve class from mollusc phylum. The main characteristic from this class is they have 2 shells, soft body with small foot, byssal gland and gills are paired. Pearl oyster can be found in tropic region and grow in clear water. Bivalves are an ideal marine organism to cultivate. These animals belong to the group of

herbivores that do not require additional food because they feed phytoplankton that naturally occur. (Mathiessen, 2001; Haws, 2002; Powell *et al.*, 2002, 2004; Fernandez *et al.*, 2006). They reproduce by releasing sperm and eggs in water column. Less than 24 hours, the fertilized eggs will develop into trochophore larvae and swim freely in the water. The larvae will metamorph into juvenile and attach to the spat. The oyster can attach, migrate and reattach by using its byssus (Haws, 2002).

The oyster eat plankton by filtering the surrounding water into filtering system and ingest small particle that contain organic matter (Dunphy *et al.*, 2006). This mechanism was happen due to coordinated small cilia movement on gill surface. The particles larger than 10 µm will expelled. The digestive system of oyster cannot break down cellulose matter.

As phytoplankton are the main food, the biochemistry composition play key role to determine the nutrition quality in phytoplankton and its function for bivalve. Biochemistry model are used to stimulate the growth and survival of bivalve larvae in various conditions (Powell *et al.*, 2002, 2004). The model shows that the increased survival rate larvae bivalve associated with low protein and high levels of lipid and carbohydrate. This composition is very important during bivalve the an attachment (Martínez-Fernández and Southgate, 2007). The larval growth is influenced by the level of carbohydrate, lipid and protein contained in phytoplankton (Fernández and Salmon, 2004; Southgate, 2006).

The aim of this research was to analyse the phytoplankton composition and its relation to water quality in pearl farming (*Pinctada maxima*) in Sekotong.

II. METHOD

2.1. Sampling Phytoplankton

Samples were taken in 2 locations Sekotong, West Nusa Tenggara. The parame-

ters that will be measured including: biological (phytoplankton), the chemical (nutrient) and physical (temperature, salinity).

Phytoplankton was taken by using 20 µm net tow. Samples were taken at the surface and depth. For surface sampling, the net plankton was towed 100 m horizontally. The water in the cod was transferred to 50 ml vial. Van Dorn was used to take the sample in 7 m deep. The water sample that was taken was the same like surface sampling. All samples were preserved by adding 0,5ml /50ml Lugol solution (Barus, 2004). Samples were taken to LIPI Mataram and ITS Surabaya for identification and documentation. Phytoplankton was identify according to Omura *et al.* (2012) and Horner (2002). The phytoplanktons were quantified using Sedgewick rafter. The density were counted using formula:

$$K = \frac{n}{m} \times \frac{s}{a} \times \frac{l}{v} \dots\dots\dots (1)$$

explanation: K= density (cells/L), n= number of counted cell in m, m= number of drop, s= sample volume, a= volume of sample, and v= volume of filtered water (m³)

Ammonia, nitrate, nitrite and phosphate concentration were measured in KKP Lombok laboratory using Lambda XLS spectrophotometer.

III. RESULT AND DISCUSSION

3.1. Phytoplankton Composition

There were 23 species of phytoplankton found in Sekotong West Nusa Tenggara (Table 1). Bacillariophyceae, Dinophyceae and Dictyochophyceae are families among phytoplankton that were found. The highest density among them was Bacillariophyceae (more than 99%).

Bacillariophyceae have wide range for its adaptation which may survive from environmental change. Most of phytoplankton in Bacillariophyceae class are euryhaline

Table 1. Phytoplankton densities in Sekotong, West Nusa Tenggara, Indonesia.

Class	Genus	Density (Cell/L)		Percentage (%)		
		Location 1	Location 2	Location 1	Location 2	
Bacillariophyceae	<i>Skeletonema</i>	814.713	484.010	83.3	83	
	<i>Chaetoceros</i>	79.872	29.620	8	3	
	<i>Bacteriastrium</i>	25.586	20.093	2.6	2	
	<i>Coscinodiscus</i>	246	410	0.03	0.04	
	<i>Eucampia</i>	164	82	0.01	0,008	
	<i>Hemialus hauckii</i>	82	82	0.008	0.008	
	<i>Guinardia</i>		82		0.008	
	<i>Navicula</i>	3.281	4.921	0.34	0.5	
	<i>Pseudonitzschia</i>	17.879	7.628	1.8	0.78	
	<i>Pleurosigma</i>	410	82	0.04	0.008	
	<i>Rhizosolenia</i>	4.511	4.347	0.46	0.4	
	<i>Thalassionema</i>	21.323	15.016	2.2	1.5	
	<i>Odontella</i>	164		0.01		
	<i>Thalassiosira</i>	7.874	11.153	0.8	1.1	
	<i>Entomoneis</i>	328	574	0.03	0.05	
	Dinophyceae	<i>Chylindrotheca</i>	411	3.034	0.04	0.3
		<i>Detonula</i>		656		0.07
<i>Protoperdinium</i>		738	574	0.08	0.05	
<i>Ceratium furca</i>		328	738	0.03	0.08	
Dictyochophyceae	<i>Gonyaulax sp</i>	<1				
	<i>Pyrophacus</i>		246		0.03	
<i>Dictyocha</i>	164	82	0.01	0.008		

which can survive in salinity 5‰-30‰ (Dwiponggo, 1976). They reproduce by means of vegetative cell division. The cells contain two part : the upper part (epitheca) and lower part (hypotheca). Epitheca will pairs with hypotheca to form a new cell. This ability makes phytoplankton abundant in nature, especially if supported by availability of nutrients. When the seawater rich of nutrients, the phytoplankton that belong to the diatom group can reproduce three times in 24 hours (Effendi, 2003). While the dinoflagellate group can only reproduce once in 24 hours with the same condition.

There were five species found in Sekotong (Bacillariophyceae) that its existence in quite abundant (more than 10^4 cell/L),

Skeletonema, *Chaetoceros*, *Bacteriastrium*, *Pseudonitzschia*, *Thalassionema* and *Thalassiosira* sp. From two locations, *Skeletonema* have the highest density with average percentage up to 83%. According to Veronica *et al.* (2014), there are three categories of phytoplankton abundance related to nutrient in water. The density of phytoplankton in less nutrient water is below 10^4 cell/L, adequate nutrient is between $10^4 - 10^7$ cell/L and high nutrient is above 10^7 cell/L. The condition where the density of phytoplankton is more than 10^7 cell/L is called blooming (Efrizal, 2001). During research this condition was not found. *Skeletonema* sp domination was particularly caused by euryhaline characteristic, which is can grow in temperature

from 3 – 30°C. This characteristic may cope the environment fluctuation (Basmi, 1999).

3.1. The Relation between Phytoplankton Composition with Pearl Farming Location

Phytoplankton play key role for pearl farming. The density and composition of phytoplankton are some of the important factors that determine the farming location. As a primary producer in food chain, phytoplankton is used as an indicator for water prolificacy.

The result showed that the quantity of the phytoplankton found in Sekotong, was abundant. They were used as food source and help the oysters grow. Phytoplankton from Bacillariophyceae, and the a including is: *Leptocylindricus* sp., *Thalassionema* sp., *Chaetoceros* sp., *Nitzschia* sp., *Rhizosolenia* sp., *Bacteriastrum* sp., and *Asterionella japonica*, generally used in pearl farming even since hatchery stage (Fukumuri *et al.*, 2007). Furthermore, the composition of food source in oyster stomach (*Pinctada fucata*) was phytoplankton (78%) and attached microalgal (22%), respectively.

Another group of phytoplankton that can produce toxin were also found in area. The densities of Dinophyceae family that can produce oxidic acid (toxin) were low. This group can be in rest stage by forming cyst and also can regrow in suitable environment (Gosling, 2003). This family have the most toxic derivate among other toxic phytoplankton group (Hamzah, 2003). The density of group in pearl farming area was 1189 cell/L blooming stage is 5000 cell/L (Basmi, 1999).

Based on the description above, the area is suitable for pearl farming. The water condition can be measured from phytoplankton composition. The composition of phytoplankton will low if the water is polluted or the nutrient is low (Handayani, 2008).

3.2. Physical and Chemical

The water temperature in both location in Sekotong were 28°C - 29°C. Accord-

ing to ministerial decision from Environmental Ministry the range is suitable for pearl oyster life. The temperature in tropical region is 26-30°C (16). Most of the marine organism in tropic region can tolerate until 20 - 30°C (Haumau, 2005).

The salinity in sampling area is 30‰. According to water standard quality the Salinity is save for oyster. Subsequently, Augusta (2012) explained that the range of salinity for pearl oyster is 15 - 35‰.

Acidity has effect on growth of marine organism. The water with high acidity can cause mortality (Haumau, 2005). The acidity number from sampling locations are 8,09 (location 1) and 7,67 (location 2). According to Augusta (2012) the acidity range in Indonesia's water is 6,0 to 8,5 while the optimum acidity for aquaculture is 7 – 8,5.

Average dissolved oxygen in locations are 4,3 ppm (location 1) and 6,3 (location 2). The standard quality of dissolved oxygen is vary (Hendrawati *et al.*, 2007) but the minimum range is between 4 – 5 ppm. Another chemical quality that must be considered for pearl farming is nitrogen concentration. The nitrite form can bond with blood and become toxic to the organism. Nitrite concentration in both location were 0,01 – 0,02 mg/l.

Phosphate concentration in water can be as indicator for pollution. In nature, they usually in organophosphate or polyphosphate form. High phosphate concentration can be caused by eutrophication that can make the growth of phytoplankton exponentially. The optimum concentration for phytoplankton growth is 0,27 – 5,51 ppm (Hendrawati *et al.*, 2007). The phosphate concentration in both locations is in optimum condition for phytoplankton growth (0,3 – 0,5 mg/l.).

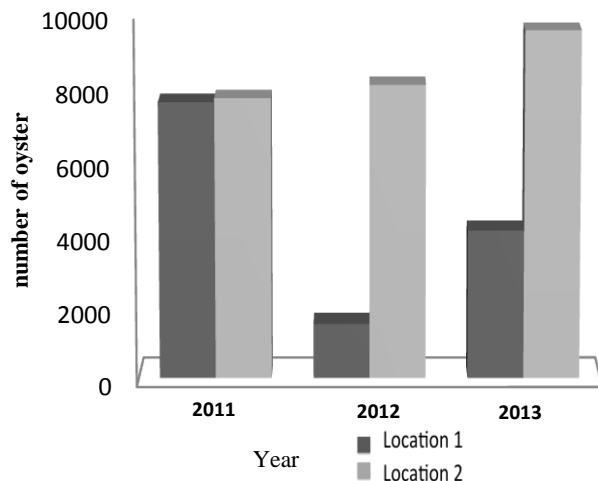
Ammonia concentration in location 1 was 0,018 mg/l (surface) and 0,21 mg/l (7 m); location 2 was 0,015 mg/l (surface) and 0,018 mg/l (7 m). The ammonia in water is mostly from organism metabolism and organic waste decomposition by bacteria (Hen-

drawati *et al.*, 2007). The standard quality of ammonia in water is 0,3 mg/l. this mean that the ammonia concentration in locations were suit for marine aquaculture including pearl farming.

3.3. Water Quality in Sekotong and the Pearl Farming Production

Phytoplankton composition in Sekotong consists of 24 species. *Skeletonema* sp., *Chaetoceros* sp., *Thalassionema* sp., and *Pseudonitschia* sp are phytoplankton species that used to feed pearl oyster. In Sekotong their density were above 10^4 cell/l. The physical and chemical parameter of water in Sekotong is accordance with the ministerial decision of Lingkungan Hidup number 51/2004.

The yield of pearl oyster farming in BBL is shown in Pic 2. The production was increasing in location 2. The production fluctuation in location 1 was particularly caused by biofouling / attached the organism.



Picture 1. Production of pearl oyster in Balai Benih Laut (BBL).

The feeding competition between oyster and biofouling such as barnacle (Isnanty and Kuniastuty, 1995) may affect the growth of oyster.

IV. SUMMARY

Bacillariophyceae was class of phytoplankton that dominant in Sekotong water. Most of the phytoplankton from this class was used to feed the oyster farming. Harmful phytoplankton from Dinophyceae class was found but in a small amount. The physical and chemical characteristic in water showed that the nutrition is adequate for phytoplankton growth and it was required for oyster viability.

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