

# **Research Article**

Check for updates



# The Antiaging Potential of Serum Formulations from *Centella asiatica*, *Curcuma longa*, *Aloe vera*, *Rosa centifolia*, and Salmon DNA on Injured Human Fibroblast Cells

Ermi Girsang<sup>1\*</sup>, Teresa Liliana Wargasetia<sup>2</sup>, Deni Rahmat<sup>3</sup>, Marisca Evalina Gondokesumo<sup>4</sup>, Mathelda Weni Harjanti<sup>2</sup>, Wahyu Widowati<sup>2</sup>, Fadhilah Haifa Zahiroh<sup>5</sup>, Zahra Qisthi Saufa<sup>6</sup>, Oktaviana Takasenserang<sup>7</sup>, Dhanar Septyawan Hadiprasetyo<sup>8</sup>

<sup>1</sup>Faculty of Medicine, Universitas Prima Indonesia, Medan 20118, North Sumatera, Indonesia

<sup>2</sup>Faculty of Medicine, Maranatha Christian University, Bandung 40164, West Java, Indonesia

<sup>3</sup>Faculty of Pharmacy, University of Pancasila, Jakarta 12640, Indonesia

<sup>4</sup>Department Biology Pharmacy, Faculty of Pharmacy, University of Surabaya, Universitas Surabaya, Surabaya 60293, Indonesia <sup>5</sup>Aretha Medika Utama, Bimolecular and Biomedical Research Center, Bandung 40163, West Java, Indonesia

<sup>6</sup>Department of Biology, School of Life Sciences and Technology, Bandung Institute of Technology, Bandung 40132, West Java, Indonesia <sup>7</sup>Department of Biology, Faculty of Mathematics and Natural Sciences, Sam Ratulangi University, Manado 95115, North Sulawesi, Indonesia

<sup>8</sup>Faculty of Pharmacy, Universitas Jenderal Achmad Yani, Cimahi 40531, West Java, Indonesia

#### ARTICLE INFO

Article history: Received October 23, 2024 Received in revised form December 17, 2024 Accepted December 20, 2024

KEYWORDS: Antiaging serum, Fibroblast cells, Gene expression, Plant Extract, Salmon DNA



Copyright (c) 2025@ author(s).

# ABSTRACT

External aging factors such as UV exposure, pollution, and lifestyle choices contribute to skin aging, resulting in deep wrinkles, fine lines, and rough skin, which can lower self-confidence. Plant extracts have been widely studied for their antiaging potential, while Salmon DNA has shown promise in stimulating collagen production. This study explores the formulation of a serum combining *Centella asiatica*, *Curcuma longa*, *Aloe vera*, *Rosa centifolia*, and Salmon DNA for its antiaging effects on injured human fibroblast cells. The serum was formulated using extracts from *C. asiatica*, *C. longa*, *A. vera*, *R. centifolia*, and Salmon DNA. Antioxidant activity was evaluated with the DPPH method, cytotoxicity using the WST-8 assay, and gene expression through qRT-PCR for COL1A1, TGF- $\beta$ 1, HYAL-1, and FGF-2. The serum exhibited weak antioxidant activity (IC<sub>50</sub> = 373.33 µg/ml) and reduced cell viability at high concentrations. Gene expression analysis revealed increased expression of COL1A1, TGF- $\beta$ 1, and FGF-2, along with reduced HYAL-1 expression in injured BJ cells. The formulated serum shows potential as an antiaging agent, promoting collagen production and reducing hyaluronidase activity.

# 1. Introduction

Aging is a normal multifactorial pattern that triggers physical alterations in the skin and connective tissue (Widowati *et al.* 2018; Zorina *et al.* 2022). These physiological changes result from intrinsic aging and cumulative extrinsic damage, including exposure to ultraviolet radiation, environmental pollution, and free radicals (Widowati *et al.* 2022).

\* Corresponding Author

These factors gradually change the shape and function of each layer of the skin, thereby changing the appearance of the skin. In intrinsic aging, the skin undergoes morphological and physiological changes such as dryness, wrinkles, and sagging, and wound healing becomes slower. Extrinsic aging causes deep wrinkles, loss of elasticity, and a rough skin surface (Yusharyahya 2021). Aging is closely related to skin regeneration and skin cell proliferation. Several key genes are important in regulating this mechanism, especially COL1A1, TGF- $\beta$ 1, HYAL-1, and FGF-2.

E-mail Address: ermigirsang@unprimdn.ac.id.

COL1A1 is known as the gene encoding the alpha-1 chain of type I collagen, a major structural element of the extracellular matrix (ECM) in the dermis. COL1A1 expression decreases with aging, causing a decrease in collagen production and contributing to skin aging (Iriyama et al. 2022). TGF-β1 is a growth factor that governs various cellular processes, such as cell proliferation, differentiation, and ECM production. It plays a crucial role in skin homeostasis, and its dysregulation has been linked with skin aging (Shin et al. 2019). HYAL-1 is a hyaluronidase enzyme that degrades hyaluronic acid (HA), a key component of the ECM. Changes in HYAL-1 expression and HA metabolism have been linked to skin aging and various age-related pathologies (Shin et al. 2019). FGF-2 is a fibroblast growth factor that promotes cell proliferation, migration, and ECM production. Its expression is important for maintaining skin homeostasis, and its downregulation has been correlated with skin aging (Hatzirodos et al. 2019). In this regard, antioxidants also play an important role in skin regeneration by modulating gene expression and creating an environment conducive to tissue regeneration. Antioxidants help neutralize excess reactive oxygen species (ROS), which can interfere with skin regeneration and contribute to skin aging (Fitzmaurice et al. 2011).

Natural cosmetics are increasingly in demand because of their positive benefits for skin health and appearance, especially through beauty treatments that utilize natural ingredients from plants and animals. These treatments are considered safer than synthetic chemical products that can cause irritation or side effects, especially for sensitive skin. Amid increasing awareness of the importance of skin health, more and more people are turning to natural cosmetics because they have minimal risk of toxicity and are in line with global trends that are more environmentally friendly. One of the most sought-after cosmetic products is serum products. Serum is a topical cosmetic with a concentrated amount of active ingredients, so it has good potential for effectiveness (Amnuaikit et al. 2022).

Salmon DNA is known to help in the formation, repair, and restructuring of tissue, which increases the formation of elastin and collagen (Lee *et al.* 2017; Sato *et al.* 2017; Sveen *et al.* 2023). Salmon DNA also contains vitamins, peptides, and antioxidants in addition to hyaluronic acid. Therefore, salmon DNA can help in skin tissue regeneration (Sato *et al.* 2017).

Salmon DNA was used in this study as the main ingredient in the formulation of antiaging serum with the addition of Centella asiatica, Curcuma longa, Aloe vera, and Rosa centifolia extracts to increase collagen and elastin synthesis, as well as improve skin structure and elasticity. Phytochemical screening revealed that the extract of C. asiatica contained alkaloids, saponins, tannins, phenolics, flavonoids, glycosides, terpenoids, and steroids (Ferdous et al. 2017). Meanwhile, turmeric extract (C. longa) contains flavonoids, polyphenols, saponins, tannins, glycosides, and secondary metabolites of alkaloids, steroids, terpenoids, and essential oils (Chanda & Ramachandra et al. 2019). Phytochemical screening of A. vera extract using 96% ethanol solvent showed that the sample contained flavonoids, phenolics, tannins, and saponins (Bista et al. 2020). The R. centifolia flower petal extract contained alkaloids, carbohydrates, flavonoids, glycosides, saponins, steroids, tannins, triterpenoids, and phenolic compounds (Nimbal et al. 2021). Overall, the bioactive components in these extracts are known to stimulate collagen production, promote cell regeneration, and reduce inflammation, so they have the potential as active ingredients in antiaging cosmetics.

Until now, there has been no research that formulates these active ingredients into serum preparations for antiaging, so the formulation and testing of its antiaging potential in vitro was carried out on injured human fibroblast cells as an antiaging model to see its cell regeneration activity. Thus, this study aims to formulate an antiaging serum and test its aging potential by measuring antioxidant parameters with 1,1-diphenyl-2-picrylhydrazyl (DPPH), cytotoxic assay, and measuring the COL1A1, TGF- $\beta$ 1, HYAL-1, and FGF-2 genes expression of in injured human fibroblast cells.

## 2. Materials and Methods

#### 2.1. Formulation of Serum

The serum product was manufactured at PT. Dizza Karya Utama, which is certified for Good Manufacturing Practices (GMP) by the Indonesian Food and Drug Authority (Indonesian FDA with the certification number PW-S.03.011.44.441.12.22-0457).

The following serum base ingredients contain water as the primary component, Na<sup>2</sup>EDTA, Carbomer (Repoly 140), Xanthan Gum (Keltrol CGSFT), Hyaluronic Acid, Glycerin KOH (Potassium Hydroxide), Phenoxyethanol BMP 800 (Methylpropanediol), Peptide Complex (Natori Peptide Complex), acetyl tetrapeptide-5 (Natori Peptide Complex), Palmitoyl tripeptide-1 (Natori Peptide Complex), Palmitoyl pentapeptide-4 (Natori Peptide Complex), Glycine max polypeptide (Natori Peptide Complex), Saccharomyces polypeptides (Natori Peptide Complex), 1,2-hexanediol (Natori Peptide Complex), d-panthenol (D-Panthenol), Allantoin (Allantoin), Vitamin E (Tocopherol), Glutathione, PEG-40 Hydrogenated Castor Oil (Sabowax), and Perfume (Fragrance).

The active ingredients of the serum can be seen in Table 1.

# 2.2. DPPH Test for Antioxidant

Serum was tested by DPPH test with serial concentrations, namely 200, 100, 50, 25, 12.5, and 6.25 µg/ml. Serum on various concentrations was added 50 µL to each well of the 96-well plate, then 200 µL DPPH (1,1-diphenyl-2 -picrylhydrazyl) solution (Sigma-Aldrich, D9132) was added to each well. The concentration of DPPH used is 0.077 mmol/L (dissolved in methanol). After that, the plate was incubated in the dark room for 30 minutes at room temperature. After incubation, the absorbance of each well was measured by a microplate reader (MultiskanTM GO Microplate Spectrophotometer, Thermo Scientific) at 517 nm wavelength. The scavenging activity was quantified using this formula (Widowati *et al.* 2018):

Scavenging activity (%) =  $(Ac - As) / Ac \times 100$ 

Ac = negative control absorbance (without sample)As = sample absorbance.

## 2.3. Cells Viability Assay

The cells used in this study are BJ fibroblast cells obtained from ATCC<sup>®</sup> CRL-2522. The cells were cultured in medium MEM with supplement complete following the method by Widowati *et al.* (2019). The cells were incubated at 5% CO<sub>2</sub> in 37°C. After cells were confluent around 70-80%, cells were counted by

Table 1. A	Active	ingredients	of	serun
------------	--------	-------------	----	-------

Table 1. Active ingredients of setuin	
Ingredients	Percentage
Centella asiatica Extract	0.5-2
Curcuma longa Extract (Turmeric)	0.5-2
Aloe vera extract	0.5-2
Rosa centifolia extract	0.5-2
Salmon DNA	0.5-2
Honey Extract	0.5-2
Niacinamide	4-6

hemocytometer and then cultured at a density of  $1 \times 10^4$  cells/well in a 96-well plate and incubated at 37°C for 24 hours, 5% CO<sub>2</sub>. After incubating, the cells medium was replaced by mediums that contained serum with serial concentrations 50, 25, 12.5, 6.25, 3.13, 1.63, and 0.82% with a volume of 100 µL in each well. After 24 hours of incubation, 10 µL WST 8 (Elabscience, E-CK-A362) was added to each well and incubated at 37°C, 5% CO<sub>2</sub> for 3 hours. Absorbance is measured by a spectrophotometer in 450 nm wavelength (Widowati *et al.* 2021; Eder *et al.* 2022).

# **2.4.** COL1A1, TGF-β1, HYAL-1, FGF-2 Genes Expression Assay

Cells were counted by hemocytometer and then cultured at a density of  $5 \times 10^5$  cells/well in a 6-well plate and incubated for 24 hours at 37°C, 5% CO<sub>2</sub>. Then, cells were wounded by making a straight line using blue tips (1 ml) in the middle of the well to create an aging model. Culture mediums were replaced by new ones containing serum with serial concentrations (4, 2, and 0.5%) and incubated for 72 hours at 37°C, 5% CO<sub>2</sub>. After that, cells were harvested to be tested for gene expression using qRT-PCR. The levels of COL1A1, TGF-β1, HYAL-1, and FGF-2 gene expression were measured from pellet cells. The total RNA isolate applied was the Direct-zol RNA Miniprep Plus Kit (Zymo, R2073), the procedure applied following the protocol from the manufacturer. The concentration and purity of RNA of each sample were measured at 260/280 nm (Table 2). A Sensi-FAST cDNA synthesis kit was used to synthesize the complementary DNA using the manufacturer's protocol. Quantitative gene expression was quantified using AriaMx 3000 Real-Time PCR System (Agilent, G8830A) with primer sequence (Table 3) and Sensi-FAST Syber NO-ROX reaction mixture. The procedure was according to the manufacturer's protocol. Real-time PCR was run for 40 cycles using GAPDH as a reference gene, and the annealing temperatures were 58°C, 59°C, 59°C, and 63°C for COL1A1, TGF-β1, HYAL-1, and FGF-2, respectively (Privandoko et al. 2024).

Table 2. Concentration and purity of RNA

Sample	Concentration (ng/µL)	Purity ( $\lambda 260/\lambda 280$ ) nm		
NC	13.60	2 258		
PC	10.24	1.345		
Serum 4%	8.88	2.345		
Serum 2%	10.56	2.154		
Serum 0.5%	9.20	2.140		

#### 2.5. Statistical Analysis

Data was analyzed using SPSS software 20.0 (SPSS Inc). For normally distributed and homogenous data, one-way ANOVA was used, followed by Tukey HSD post-hoc test. The data's significance level is P-value < 0.05. The data was visualized using GraphPad Prism (version 9.2.0.332), which displays mean±standard deviation in histograms (Widowati *et al.* 2021).

## 3. Results

# **3.1. Effect of Formulated Serum on DPPH** Scavenging Activity

The antioxidant activity of serum was analyzed using the DPPH method. The results showed the antioxidant activity of the serum. DPPH inhibition is in line with the reduction of serum concentration. The serum showed concentration-dependent antioxidant activity (Figure 1) with a median Inhibitory Concentration (IC<sub>50</sub>) 373.33  $\mu$ g/ml (Table 4).

# **3.2. Effect of Formulated Serum on Fibroblast** Cell Viability

The results of the study showed that the serum from the formulation had toxic levels to cells if in high concentrations. This data can be seen in Figure 2A. The higher the concentration of serum given, the lower the cell viability. A serum concentration of 0.82% had exhibited viability similar to the negative control. Inversely proportional to serum on BJ cell inhibition (Figure 2B), the lower the concentration of serum given, the lower the inhibition of cell viability. For further research, the safest concentration was used with viability above 80%.

# **3.3.** Effect of Formulated Serum Toward FGF-2, HYAL-1, COL1A1 and TGFB-1 Gene Expression in Injured-Skin Fibroblast

Based on the results obtained, serum had a regulatory effect on gene expression related to cell

Table 3. Primer sequence

regeneration in injured BJ cells. The scratch results on BJ cells can increase the expression of FGF-2 and HYAL-1 genes while decreasing COL1A1 and TGF- $\beta$ 1 (Figure 3). In this case, serum can improve the genes related to regeneration regulation. The results showed that FGF-2 gene expression in BJ cells as an aging cells model treated with serum increased relatively compared to the positive control. The most effective concentration in enhancing FGF-2 gene expression was 0.82%. Serum also decreased HYAL1 gene expression in injured BJ cells. Furthermore, the data showed that serum could increase COL1A1 and TGF- $\beta$ 1 gene expression in aging cells compared to the positive control. The data shown is the average



Figure 1. Effect of serum concentrations in DPPH scavenging activity. I: serum 6.25  $\mu$ g/ml, II: serum 12.5  $\mu$ g/ml III: serum 25  $\mu$ g/ml, IV: serum 50  $\mu$ g/ml, VI: serum 100  $\mu$ g/ml, VI: serum 200  $\mu$ g/ml, Data are represented in mean  $\pm$  standard deviation. The different letters in the graph show significant differences among various serum concentrations based on the Tukey HSD Post Hoc Test P<0.05

Table 4. The antioxidant activity (IC<sub>50</sub>) values of formulated serum

Assay	Linear Equation	$IC_{50}$ value (µg/ml)
DPPH	y = 0,0914x + 15,878	373.33

	1				
Gene symbols	Primer sequence (5' to 3')	Product size (bp)	Annealing (°C)	Cycle	References
COL1A1	F: GAATTCGGCTTCGACGTTGG	127	58	40	NM_000088.4
TGF-β1	F: GACTTTTCCCCAGACCTCGG	135	59	40	NM_000660.7
HYAL-1	R: ATAGGGGATCTGTGGCAGGT F: GCCCTTCATCCTGAACGTGA	138	59	40	NM_153281.2
FGF-2	R: AGCTGGATGGAGAAACTGGC F: GAGCCCAGGAGTTCAAGACC	93	63	40	NM_002006.6
GAPDH	R: GAGACCACATGTACACGCCA	172	58	40	NM 001289745.3
UAI DII	R: GAAGATGGTGATGGGATTTC	1/2	20	10	_



Figure 2. Effect of serum concentrations on the viability of BJ cells as measured based on (A) viability and (B) inhibitory value. I: Negative control II: serum 50 %, III: serum 25 % IV: serum 12.5 %, V: serum 6.25 %, VI: serum 3.13 %, VII: serum 1.63 %, VII: serum 0.82 %, Data are represented in mean ± standard deviation. The different letters in the graph show significant differences among various serum concentrations based on the Tukey HSD Post Hoc Test P<0.05</p>



Figure 3. Effect of serum concentrations toward genes expression of (A) COL1A1, (B) TGF-β1, (C) HYAL-1, and (D) FGF-2. I: Negative control II: Positive control, III: Serum concentration 4%, IV: Serum concentration 2%, V: Serum concentration 0.5%, Data are presented in mean ± standard deviation. Different letters in each bar showed significant differences between treatments based on the Tukey HSD Post Hoc Test P<0.05</p>

of three replications where the lower the serum concentration is given, the higher the repair effect.

#### 4. Discussion

Skin aging is a global concern that many women complain about. Preventing skin aging can be achieved by using skincare products with active antiaging ingredients. Serum is a widely used skincare formulation because it contains active ingredients at high concentrations, which allows for faster absorption into the skin, and its low viscosity makes it easy to spread over the skin's surface (Amnuaikit et al. 2022). In this study, the serum is formulated with active ingredients that have been proven in many studies to be beneficial for antiaging. R. centifolia extract has anti-inflammatory, antibacterial, anti-allergic, and antiaging properties. C. asiatica is known to help wound healing and skin inflammation through reepithelialization and stimulation of collagen synthesis. Its bioactive compounds, including madecassic acid, madecassoside, and asiatic acid, also provide antiinflammatory, antioxidant, and antimicrobial properties (Yasurin et al. 2016). According to Kim et al. (2019), turmeric leaf extract contains antioxidant components like phenolic and flavonoid compounds that can scavenge ROS. Turmeric leaf water extract is known to have a high total phenolic content  $(2.741\pm0.099)$ mg GAE/g) and flavonoid content (4.776±0.010 mg QCE/g). In this study, salmon DNA was also added to the serum formulation to support tissue formation, repair, and remodeling, as well as to enhance collagen and elastin synthesis, improving skin manifestations including sagging, loss of firmness, dryness, dullness, and uneven skin tone (Lee et al. 2017; Sato et al. 2017 Sveen et al. 2023).

Antioxidant assay was conducted using the DPPH method to test the activity of the serum's active ingredients. DPPH was used as a free radical source to measure antioxidant activity. The DPPH test results showed that the higher the serum concentration, the greater the DPPH scavenging activity (Figure 1). The antioxidant ability of a compound is classified into several groups. IC<sub>50</sub> values < 50 µg/ml are 'very strong', IC<sub>50</sub> values of 50-100 µg/ml are 'strong', IC<sub>50</sub> values of 250-500 µg/ml are 'weak', and IC<sub>50</sub> values of more than 500 µg/ml are inactive antioxidants. The serum showed antioxidant activity with IC<sub>50</sub> 373.33 µg/ml and was classified as a weak antioxidant (Kusumawati

*et al.* 2021). These data indicate that the serum has antioxidant activity due to its active ingredients, such as *C. longa* and *C. asiatica*. Based on DPPH, ABTS,  $H_2O_2$ , NO, and FRAP tests, *C. longa* extract is known to have high antioxidant activity (Laksmitawati *et al.* 2022). *C. asiatica* extract also exhibits high antioxidant activity in DPPH scavenging tests (Buranasudja *et al.* 2021). *C. asiatica* is known to contain a high amount of flavonoids such as apigenin, catechin, kaempferol, rutin, quercetin, and naringin, making this plant a rich source of antioxidants (Taghizadeh & Jalili 2023). With these active ingredients, the serum provides excellent antioxidant effects.

The cell viability test results show that the serum is safe to use at low concentrations (Figure 2). This can be seen in Figure 2A, where the lower the serum concentration, the higher the cell viability. The serum concentration of 0.82% showed cell viability similar to the negative control. This served as the basis for selecting the concentration for the next study, qRT-PCR.

The effects of serum on FGF-2 gene expression in injured BJ cells showed that FGF2 gene expression increased depending on the concentration applied. Low-concentration serum can increase FGF-2 gene expression more significantly in injured BJ cells. FGF2 is a growth factor that plays an important role in tissue regeneration and repair. It is associated with aging due to its ability to influence cell proliferation, angiogenesis, and differentiation, which can contribute to decreased tissue function with age (Farooq *et al.* 2021). The presence of active ingredients like aloin from *A. vera* may be responsible, as studies have shown that *A. vera* gel extract can increase FGF-2 gene expression (Razi *et al.* 2021).

HYAL-1 is a hyaluronidase enzyme responsible for degrading hyaluronic acid in the skin, and its activity decreases with age, leading to lower hyaluronic acid levels, contributing to the loss of moisture and elasticity, and accelerating signs of aging (Abatangelo *et al.* 2020). Scratch induction increases HYAL-1 gene expression, which can lead to skin aging. The serum significantly reduced HYAL-1 gene expression in injured BJ cells. This may be due to the active ingredients in the serum, as previous studies have shown that *A. vera* extract can modulate HYAL-1 expression in HaCaT cells (Razia *et al.* 2021).

The COL1A1 gene plays a role in collagen synthesis, a key component of the skin's extracellular matrix. Based on the results, the tested serum increased COL1A1 gene expression compared to the positive control. The serum also enhanced TGF- $\beta$ 1 gene expression compared to the positive control. Several studies have shown that TGF- $\beta$ 1 expression plays an essential role in regulating cell growth and collagen formation (Ansary *et al.* 2021). The proposed mechanism of serum as antiaging can be seen in Figure 4.

The diagram illustrates a proposed mechanism for the antiaging effects of a serum formulation containing active ingredients from C. asiatica (asiaticoside), C. longa (curcumin), A. vera (aloin), R. centifolia (quercetin), and salmon DNA. The pathway suggests that these ingredients may influence key aspects of cellular aging, including telomere shortening, oxidative stress, cellular senescence, and chronic inflammation (López-Otín et al. 2013). The active compounds appear to modulate several important molecular pathways. For instance, the upregulation of COL1A1 could enhance collagen production, potentially improving skin firmness and elasticity (Quan et al. 2010). The increased expression of TGF-β1 may promote collagen synthesis and tissue repair, which are crucial for maintaining skin structure and function (Pakyari et al. 2013). Additionally, the stimulation of FGF-2 could boost fibroblast proliferation and tissue regeneration, contributing to skin rejuvenation (Xie et al. 2015).

The downregulation of HYAL-1 suggests a potential mechanism for improving skin hydration by reducing

hyaluronic acid degradation (Papakonstantinou et al. 2012). This multi-faceted approach targeting various aspects of skin aging aligns with the current understanding of the complex nature of the aging process and the need for comprehensive interventions (Ganceviciene et al. 2012). The focus on oxidative stress in the pathway is particularly noteworthy, as it is a well-established contributor to skin aging (Rinnerthaler et al. 2015). The antioxidant properties of ingredients like curcumin and quercetin may play a significant role in combating oxidative damage and its downstream effects on cellular senescence and inflammation (Boots et al. 2008; Altundağ et al. 2021). While this mechanism provides a theoretical framework for the antiaging potential of the serum formulation, further experimental validation would be necessary to confirm these pathways in human fibroblasts and skin tissue.

## Acknowledgements

This research was funded by Pendanaan Riset dan Inovasi Indonesia Maju (RIIM) Badan Riset Inovasi Nasional (BRIN) (III/IV/KS/2023). Laboratory facilities and research methodology were provided by Aretha Medika Utama, Biomolecular and Biomedical Research Center, Bandung, Indonesia. We are grateful to PT Dizza Karya Utama for processing the formulated serum.



Figure 4. Proposed mechanism formulated serum on aging cell



#### References

- Abatangelo, G., Vindigni, V., Avruscio, G., Pandis, L., Brun, P., 2020. Hyaluronic acid: Redefining its role. *Cells.* 9, 1743. https://doi.org/10.3390/cells9071743
- Altundağ, E.M., Özbilenler, C., Ustürk, S., Kerküklü, N.R., Afshani, M., Yilmaz, E., 2021. Metal-based curcumin and quercetin complexes: Cell viability, ROS production and antioxidant activity. *J. Mol. Struct.* 1245, 131107. https:// doi.org/10.1016/j.molstruc.2021.131107
- Amnuaikit, T., Shankar, R., Benjakul, S., 2022. Hydrolyzed fish collagen serum from by-product of food industry: Cosmetic product formulation and facial skin evaluation. *Sustainability* 14, 16553. https://doi.org/10.3390/su142416553
- Ansary, T.M., Hossain, M.R., Kamiya, K., Komine, M., Ohtsuki, M., 2021. Inflammatory molecules associated with ultraviolet radiation-mediated skin aging. *Int. J. Mol. Sci.* 22, 3974. https://doi.org/10.3390/ijms22083974
- Bista, R., Ghimire, A., Subedi, S., 2020. Phytochemicals and antioxidant activities of *Aloe vera (Aloe barbadensis)*. *J Nut Sci Heal Diet.* 1, 25-36. https://doi.org/10.47890/ JNSHD/2020/RBista/10243803
- Boots, A.W., Haenen, G.R., Bast, A., 2008. Health effects of quercetin: From antioxidant to nutraceutical. *Eur. J. Pharmacol.* 585, 325-337. https://doi.org/10.1016/j. ejphar.2008.03.008
- Buranasudja, V., Rani, D., Malla, A., Kobtrakul, K., Vimolmangkang, S., 2021. Insights into antioxidant activities and anti-skinaging potential of callus extract from *Centella asiatica* (L.). *Sci. Rep.* 11, 13459. https://doi.org/10.1038/s41598-021-92958-7
- Chanda, S., Ramachandra, T.V., 2019. Phytochemical and pharmacological importance of turmeric (*Curcuma longa*): A review. *Res. Rev. J. Pharmacol.* 9, 16-23.
- Eder, K.M., Marzi, A., Wågbø, A.M., Vermeulen, J.P., de la Fonteyne-Blankestijn, L.J.J., Rösslein, M., Ossig, R., Klinkenberg, G., Vandebriel, R.J., Schnekenburger, J., 2022. Standardization of an in vitro assay matrix to assess cytotoxicity of organic nanocarriers: A pilot interlaboratory comparison. *Drug Deliv. Transl. Res.* 12, 2187-2206. https://doi.org/10.1007/ s13346-022-01203-9
- Farooq, M., Khan, A.W., Kim, M.S., Choi, S., 2021. The role of fibroblast growth factor (FGF) signaling in tissue repair and regeneration. *Cells.* 10, 3242. https://doi.org/10.3390/ cells10113242
- Ferdous, N., Rahman, M., Alamgir, A.N.M., 2017. Investigation on phytochemical, cytotoxic and antimicrobial properties of ethanolic extracts of *Centella asiatica* (L.) Urban. J. Med. Plants Stud. 5, 187-188.
- Fitzmaurice, S.D., Sivamani, R.K., Isseroff, R.R., 2011. Antioxidant therapies for wound healing. Skin Pharmacol. *Physiol*. 24, 113-122. https://doi.org/10.1159/000322643
- Ganceviciene, R., Liakou, A.I., Theodoridis, A., Makrantonaki, E., Zouboulis, C.C., 2012. Skin antiaging strategies. *Dermatoendocrinol.* 4, 308-319. https://doi.org/10.4161/ derm.22804

- Hatzirodos, N., Hummitzsch, K., Irving-Rodgers, H.F., Breen, J., Perry, V.E.A., Anderson, R.A., Rodgers, R.J., 2019. Transcript abundance of stromal and thecal cell related genes during bovine ovarian development. *PLoS One.* 14, e0213575. https://doi.org/10.1371/journal.pone.0213575
- Iriyama S., Ogura Y., Nishikawa S., Hosoi J., Amano S., 2022. Regeneration of collagen fibrils at the papillary dermis by reconstructing basement membrane at the dermal-epidermal junction. *Sci Rep.* 12, 795. https://doi.org/10.1038/s41598-022-04856-1
- Kim, S., Ko, S.C., Kim, Y.S., Ha, S.K., Park, H.Y., Park, Y., Lee, S.H., 2019. Determination of *Curcuma longa* L. (Turmeric) leaf extraction conditions using response surface methodology to optimize extraction yield and antioxidant content. *J. Food Qual.* 2019, 7575206. https:// doi.org/10.1155/2019/7575206
- Kusumawati, A.H., Farhamzah, F., Alkandahri, M.Y., Sadino, A., Agustina, L.S., Apriana, S.D., 2021. Antioxidant activity and sun protection factor of black glutinous rice (*Oryza* sativa var. glutinosa). Trop J Nat Prod Res. 5, 1958-1961. https://doi.org/10.26538/tjnpr/v5i11.11
- Laksmitawati, D.R., Pratami, D.K., Widowati, W., Kusuma, H.S.W., Wijayanti, C.R., Wahyuni, C.D., Rizal, R., 2022. Antioxidant properties of *Curcuma longa* L. and *Curcuma xanthorriza* rhizomes. *SciTePress.* 1, 104-111. https://doi. org/10.5220/0010745300003113
- Lee, Y., Dugansani, S.R., Jeon, S.H., Hwang, S.H., Kim, J.H., Park, S.H., Jeong, J.H., 2017. Drug-delivery system based on salmon DNA nano-and micro-scale structures. *Sci. Rep.* 7, 9724. https://doi.org/10.1038/s41598-017-09904-9
- López-Otín, C., Blasco, M.A., Partridge, L., Serrano, M., Kroemer, G., 2013. The hallmarks of aging. *Cell*. 153, 1194-1217. https://doi.org/10.1016/j.cell.2013.05.039
- Nimbal, S.K., Gadad, P.C., Koti, B.C., 2021. Effect of ethanolic extract of *Rosa centifolia* against doxorubicin induced nephrotoxicity in albino rats. *J. Ayurveda Integr. Med.* 12, 657-662. https://doi.org/10.1016/j.jaim.2021.07.020
- Pakyari, M., Farrokhi, A., Maharlooei, M.K., Ghahary, A., 2013. Critical role of transforming growth factor beta in different phases of wound healing. *Adv. Wound Care 2*, 215-224. https://doi.org/10.1089/wound.2012.0406
- Papakonstantinou, E., Roth, M., Karakiulakis, G., 2012. Hyaluronic acid: A key molecule in skin aging. *Dermatoendocrinol.* 4, 253-258. https://doi.org/10.4161/derm.21923
- Priyandoko, D., Widowati, W., Lenny, L., Novianti, S., Revika, R., Kusuma, H.S.W., Sholihah, I.A., 2024. Green tea extract reduced lipopolysaccharide-induced inflammation in L2 cells as acute respiratory distress syndrome model through genes and cytokine pro-inflammatory. *Avicenna J. Med. Biotechnol.* 16, 57-65. https://doi.org/10.18502/ajmb. v16i1.14172
- Quan, T., Shao, Y., He, T., Voorhees, J.J., Fisher, G.J., 2010. Reduced expression of connective tissue growth factor (CTGF/CCN2) mediates collagen loss in chronologically aged human skin. J. Invest. Dermatol. 130, 415-424. https:// doi.org/10.1038/jid.2009.224

- Razi, F., Hardianto, A., Riawan, L., Priosoeryanto, B.P. 2021. Expression of fibroblast growth factor-2 after application of the Queen's crepe-myrtle leaf (*Lagerstroemia speciosa*) and *Aloe vera* extract gel in the wound healing process of hyperglycemic. *Synthesis.* 8, 9. https://doi.org/10.24198/ pjd.vol33no1.21276
- Razia, S., Park, H., Shin, E., Shim, K.S., Cho, E., Kim, S.Y., 2021. Effects of *Aloe vera* flower extract and its active constituent isoorientin on skin moisturization via regulating involucrin expression: In vitro and molecular docking studies. *Molecules*. 26, 2626. https://doi.org/10.3390/ molecules26092626
- Rinnerthaler, M., Bischof, J., Streubel, M.K., Trost, A., Richter, K., 2015. Oxidative stress in aging human skin. *Biomolecules*. 5, 545-589. https://doi.org/10.3390/biom5020545
- Sato, A., Kajiya, H., Mori, N., Sato, H., Fukushima, T., Kido, H., Ohno, J., 2017. Salmon DNA accelerates bone regeneration by inducing osteoblast migration. *PLoS One.* 12, e0169522. https://doi.org/10.1371/journal.pone.0169522
- Shin, J.W., Kwon, S.H., Choi, J.Y., Na, J.I., Huh, C.H., Choi, H.R., Park, K.C., 2019. Molecular mechanisms of dermal aging and antiaging approaches. *Int J Mol Sci.* 20, 2126. https:// doi.org/10.3390/ijms20092126
- Shin, J.S., Han, H.S., Lee, S.B., Myung, D.B., Lee, K., Lee, S.H., Kim, H.J., Lee, K.T., 2019. Chemical constituents from leaves of *Hydrangea serrata* and their anti-photoaging effects on UVB-irradiated human fibroblasts. *Biol Pharm Bull.* 42, 424-431. https://doi.org/10.1248/bpb.b18-00742
- Sveen, L.R., Robinson, N., Krasnov, A., Daniels, R.R., Vaadal, M., Karlsen, C., Tengs, T., 2023. Transcriptomic landscape of Atlantic salmon (*Salmo salar L.*) skin. G3: Genes, Genomes, Genet. 13, jkad215. https://doi.org/10.1093/ g3journal/jkad215
- Taghizadeh, M., Jalili, S., 2023. Phytochemical content, antioxidant properties, and antibacterial activities of *Centella asiatica* L. *Nat. Prod. Res.* 1-6.
- Widowati, W., Wargasetia, T.L., Afifah, E., Mozef, T., Kusuma, H.S.W., Nufus, H., Arumwardana, S., Amalia, A., Rizal, R., 2018. Antioxidant and antidiabetic potential of *Curcuma longa* and its compounds.

- Widowati, W., Gunanegara, R.F., Rizal, R., Widodo, W.S., Amalia, A., Wibowo, S.H.B., Handono, K., Marlina, M., Lister, I.N.E., Chiuman, L., 2019. Comparative analysis of Wharton's Jelly mesenchymal stem cell (WJ-MSCs) isolated using explant and enzymatic methods. J. Phys. Conf. Ser. 1374, 012024. https://doi.org/10.1088/1742-6596/1374/1/012024
- Widowati, W., Jasaputra, D.K., Kusuma, H.S.W., Rizal, R., Laksmitawati, D.R., Artie, D.S., Sholihah, I. A., Yusepany, D.T., Lestari, R., Handayani, R. A.S., Arumwardana, S., Murti, H., Bachtiar, I., Subangkit, M., 2021. Hypoxic and normoxic-human Wharton's Jelly mesenchymal stem cellfree lysate for anticancer therapies. *Walailak J. Sci. Technol.* 18, Article 9. https://doi.org/10.48048/wjst.2021.9270
- Widowati, W., Tjokropranoto, R., Damayanti, C., Kusuma, H.S.W., Handayani, T., Rizal, R. 2022. Potential of black tea (*Camellia Sinensis* (L.) O. Kuntze) extract as antioxidant and skin antiaging. *In: Proc. 1st Int. Conf. Emerging Issues Technol., Eng., and Sci,* Vol. 1. pp. 65-73.
- Xie, J., Bian, H., Qi, S., Xu, Y., Tang, J., Li, T., Liu, X., 2015. Effects of basic fibroblast growth factor on the expression of extracellular matrix and matrix metalloproteinase-1 in wound healing. *Clin. Exp. Dermatol.* 40, 173-179.
- Yasurin, P., Sriariyanun, M., Phusantisampan, T., 2016. The bioavailability activity of *Centella asiatica*. *Appl. Sci. Eng. Prog.* 9, 1-9. https://doi.org/10.14416/j.ijast.2015.11.001
- Yusharyahya, S.N., 2021. Mechanism of skin aging as a basis for prevention and treatment of aging skin: Mechanism of skin aging. Indones. *Med. eJ*. 150-150. https://doi.org/10.23886/ ejki.9.49.150
- Zorina, A., Zorin, V., Kudlay, D., Kopnin, P., 2022. Age-related changes in the fibroblastic differon of the dermis: Role in skin aging. *Int. J. Mol. Sci.* 23, 6135. https://doi.org/10.3390/ ijms23116135