

## Dynamics of intraerythrocytic parasite infections in the Java common palm civet (*Paradoxurus hermaphroditus*)

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**ABSTRACT:** Over seven weeks, this study investigated intraerythrocytic parasitic infections in the Java common palm civet (*Paradoxurus hermaphroditus*). Blood samples were collected immediately after the civet had been brought from its natural habitat. Following the 30-day adaptation period, additional samples were collected weekly. Our findings revealed infections with *Babesia* sp., *Theileria* sp., and *Anaplasma* sp., with infection rates being the highest for *Anaplasma* sp. ( $0.35 \pm 0.05\%$ ), followed by *Theileria* sp. ( $0.12 \pm 0.05\%$ ), and *Babesia* sp. ( $0.03 \pm 0.02\%$ ). Throughout the study period, the infection rates of *Babesia* sp. and *Anaplasma* sp. decreased, whereas *Theileria* sp. showed an increasing trend until the conclusion of the study.

### Keywords:

*Anaplasma* sp., *Babesia* sp., *Theileria* sp., common palm civet

### ■ INTRODUCTION

The civet (*Paradoxurus hermaphroditus*), a member of the family Viverridae, is a mammal with significant ecological and economic contributions. This genus includes four distinct species: the common palm civet (*Paradoxurus hermaphroditus*), brown palm civet (*Paradoxurus jerdoni*), golden palm civet (*Paradoxurus zeylonensis*), and Mentawai palm civet (*Paradoxurus lignicolor*) (Riffel *et al.* 1989). Civets are critical in forest ecosystems as seed dispersers that facilitate forest rejuvenation. Although their digestive system is not fully efficient, it allows them to excrete hard grains such as coffee beans that are largely undigested, thus spreading viable seeds across the forest floor (Mudappa *et al.* 2010). Beyond their ecological value, civets also have considerable economic importance.

Coffee beans collected from civet droppings are highly valued in the global market as premium-quality coffee (Cahill 2020). Moreover, the perfume industry benefits significantly from the extraction of civet musk, a substance derived from hormones used to produce luxury fragrances (Taye 2009). However, these economic benefits have led to issues concerning animal welfare and illegal trading, necessitating rigorous regulation and monitoring (Lewis-Whelan *et al.* 2024; Nijman *et al.* 2024).

Regarding health, civets are susceptible to various endoparasites, including blood parasites such as protozoa. Commonly found in both domestic and wild animals, *Babesia* sp. and *Theileria* sp. (Penzhorn 2006), and *Anaplasma* sp.

(Dyachenko *et al.* 2012), are also concerns for civets. A study by Orkun and Karaer (2017) highlighted the presence of *Babesia vulpes* in foxes and identified multiple Babesian species such as *B. rossi*, *B. crassa*, and *B. occultans* in ticks collected from wild boars and hares. Despite these findings, the prevalence of these parasites in Indonesian civets remains unreported, underscoring the need for comprehensive health assessments to identify and manage diseases in these animals.

### ■ MATERIALS AND METHODS

Upon acquisition from its natural habitat, blood samples were collected from the civet the next day. The civet underwent a 30-day adaptation period to acclimate to the new environment, after which another blood sample was collected. Subsequently, two more samples were taken at seven-day intervals. Each blood sample was carefully processed, and a drop of blood was spread into a thin smear on a slide, fixed with methanol to preserve cellular structures, and then stained with Giemsa dye for 30 min (Figure 1). Excess dye was washed away under running water to ensure that only the stained cellular components remained, and the slide was air-dried (Aroon *et al.* 2009).

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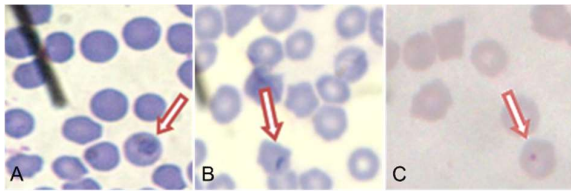


Figure 1. Blood parasites identified in civet blood smears, indicated by arrows: (A) *Babesia* sp., (B) *Theileria* sp., and (C) *Anaplasma* sp.

For microscopic examination, stained blood smears were examined under a microscope using an objective magnification of 100x and an ocular magnification of 10x. This setup facilitated detailed observation of blood parasites. The prevalence of parasites was quantified by counting the parasites per thousand red blood cells (RBCs) and expressing this count as a percentage using the formula (Lehtinen *et al.* 2008):

$$\text{Number of parasites per 1000 RBCs} \times 100\% \dots\dots\dots (1)$$

## RESULTS AND DISCUSSION

The extent and severity of parasitic infections within the red blood cells were quantified as the percentage of parasitaemia observed. Table 1 presents the average parasitaemia percentages across different time points for civets infected with *Babesia* sp., *Theileria* sp., and *Anaplasma* sp., each with varying degrees of infectivity. Notably, *Anaplasma* sp. infections exhibited the highest average parasitaemia rates, particularly on days 1 and 30, with a peak value of 0.38%.

Interestingly, while the parasitaemia levels for *Babesia* sp. remained consistent throughout the study period, from days 1 to 44, *Theileria* sp. demonstrated a notable increase in infection rates on days 30 and 37. This pattern suggests that different dynamics and possibly different immune responses are triggered by each parasite. The pronounced prevalence of *Anaplasma* sp. may be attributed to the diverse array of vectors involved in its transmission compared with the other two parasites. *Anaplasma* sp. can be spread by an extensive variety of vectors, including 20 species of ticks, tabanus flies, mosquitoes, and stable flies, providing multiple pathways for its transmission and increasing the likelihood of infection (Ashadi & Handayani 1992, Hornok *et al.* 2008).

## CONCLUSION

In these civets, primary parasitic infections of red blood cells are caused by protozoa (*Babesia* sp. and *Theileria* sp.) and rickettsia (*Anaplasma* sp.). *Anaplasma* sp. showed the highest parasitaemia levels, indicating a stronger infective capacity than *Babesia* sp. and *Theileria* sp.

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Table 1 Percentage of parasitemia for *Babesia* sp., *Theileria* sp., and *Anaplasma* sp. in civets over a 44-day acclimatization period.

Blood parasite	Blood sampling (Day), parasitemia (%)			
	1	30	37	44
<i>Babesia</i> sp.	0.03±0.08 <sup>b</sup>	0.05±0.05 <sup>b</sup>	0.02±0.04 <sup>b</sup>	0.00±0.00 <sup>b</sup>
<i>Theileria</i> sp.	0.05±0.08 <sup>b</sup>	0.12±0.19 <sup>b</sup>	0.12±0.13 <sup>b</sup>	0.17±0.10 <sup>b</sup>
<i>Anaplasma</i> sp.	0.38±0.17 <sup>a</sup>	0.38±0.10 <sup>a</sup>	0.35±0.16 <sup>a</sup>	0.27±0.18 <sup>a</sup>

Note: Different superscript letters in the same column indicate significant differences at the confidence level of 95% ( $p < 0.05$ ).

## REFERENCES

- Aroon S, Artchawakom T, Hill JG, Kupittayanant S, Thane N. 2009. Ectoparasites of the common palm civet (*Paradoxurus hermaphroditus*) at Sakaerat Environmental Research Station, Thailand. *Suranaree Journal of Science and Technology*. 16(4):277–281.
- Ashadi G, Handayani SU. 1992. *Protozoologi Veteriner I*. Bogor: IPB.
- Cahill C. 2020. [Un] becoming a resource: Translating the nature of civets in Indonesia. *Ethnos*. 85(1):100–117.
- Dyachenko V, Pantchev N, Balzer HJ, Meyersen A, Straubinger RK. 2012. First case of *Anaplasma platys* infection in a dog from Croatia. *Parasites & Vectors*. 5(49):1–7.
- Hornok S, Földvári G, Elek V, Naranjo V, Farkas R, de la Fuente J. 2008. Molecular identification of *Anaplasma marginale* and rickettsial endosymbionts in blood-sucking flies (Diptera: Tabanidae, Muscidae) and hard ticks (Acari: Ixodidae). *Veterinary Parasitology*. 154(3–4):354–359.
- Lehtinen LE, Birkenheuer AJ, Droleskey RE, Holman PJ. 2008. In vitro cultivation of a newly recognized *Babesia* sp. in dogs in North Carolina. *Veterinary Parasitology*. 151(2–4):150–157.
- Lewis-Whelan B, Ardiansyah A, Roberts PD, Nijman V, Damianou E, Morcatty TQ, Birot H, Imron MA, Nekaris KA. 2024. Welfare and Management of Civets in Civet Coffee Tourism Plantations. *Journal of Applied Animal Welfare Science*. 27(3):561–574.
- Mudappa D, Kumar A, Chellam R. 2010. Diet and fruit choice of the brown palm civet *Paradoxurus jerdoni*, a viverrid endemic to the Western Ghats rainforest, India. *Tropical Conservation Science*. 3(3):282–300.
- Nijman V, Abdullah A, Adinda E, Ardiansyah A, Campera M, Chavez J, Dewi T, Hedger K, Imron MA, Shepherd CR, Sukmadewi DK. 2024. Indonesia's sustainable development goals in relation to curbing and monitoring the illegal wildlife trade. *Sustainable Development*. 32:5393–5403.
- Orkun Ö, Karaer Z. 2017. Molecular characterization of *Babesia* species in wild animals and their ticks in Turkey. *Infection, Genetics and Evolution*. 55:8–13.
- Penzhorn BL. 2006. Babesiosis of wild carnivores and ungulates. *Veterinary Parasitology*. 138(1–2):11–21.
- Riffel M, Van Rompaey H, Schreiber A, Wirth R. 1989. Weasels, civets, mongooses, and their relatives: an action plan for the conservation of mustelids and viverrids. IUCN: International Union for Conservation of Nature. Association of Zoos and Aquariums, US, International Fur Trade Federation, IUCN Species Survival Commission (SSC), Mustelid and Viverrid Specialist Group, WWF, Zoologische Gesellschaft, DE. Retrieved from <https://coilink.org/20.500.12592/bkjq2d> on 27 Dec 2024.
- Taye T. 2009. The African Civet Cat (*Viverra civetta*) and its life supporting role in the livelihood of smallholder farmers in Ethiopia. Conference on international research on food security, natural resource management and rural development, University of Hamburg. p 1–4