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# Mineral and Vitamin B Contents of Sapudi and Merino-cross Meat

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### ABSTRACT

Lamb is important in providing balanced and healthy nutrition as a source of protein, fat, and essential micronutrients such as iron, zinc, and vitamin B complex. The mineral and vitamin content of meat from local breeds needs to be evaluated as a portrait of the genetic quality data of sheep and as a reference for developing sheep that produce healthy and high-quality meat. This study aimed to analyze the mineral and vitamin B contents of Sapudi sheep and Merino cross sheep meat from the *longissimus dorsi* area to invent genetic quality data for the meat of these two sheep. Three Sapudi and three Merino cross sheep were kept under similar conditions for two months. Meat collection from the *longissimus dorsi* muscle was carried out to analyze vitamin B and mineral content. A *t*-test was then performed to determine meat quality. The results showed that the mineral content of Mn and Cu in Sapudi sheep meat was lower than that in Merino cross meat. The vitamin B2 content in Sapudi sheep (0.11 mg/100 g) was lower than that of Merino cross (0.20 mg/100 g). In comparison, the vitamin B3 content of Sapudi sheep meat was higher (0.51 mg/100 g) than Cross-merino (0.40 mg/100 g). This research concludes that breeding influences nutrient content, and Merino crossbreed sheep are superior to Sapudi sheep in terms of vitamin B and mineral content.

Keywords: genetic resource, Indonesian local lamb, meat quality, mineral, vitamin

### INTRODUCTION

Lamb meat is one of popular source of animal protein in Indonesians. In East Java, smallholder breeders and medium-sized corporations raise a variety of sheep extensively. The sheep agriculture company mainly tries to produce males for sale during gurbani (sacrifice), ageega, and to meet daily trader demand (satay and processed lamb meat). Lamb meat also helps to maintain balance and good nutrition by providing protein, fat, and critical micronutrients including iron, zinc, and vitamin B12 (Ekmekcioglu et al. 2017). Lamb meat can also help prevent malnutrition and improve food security in impoverished countries (Mlambo and Mapiye 2015). Consumers who are aware of good consumption patterns are more likely to choose healthy food sources, particularly red meat. Meat is an essential food source that also contains minerals. Lamb meat contains little calcium but is high in potassium, sodium, zinc, iron, and phosphorus (Belhaj et al. 2021). The level of complex B vitamins, which humans require, is another indicator of chemical quality. According to Rekanović et al. (2019), meat products are a rich source of macro- and micronutrients, with cooked meat having a higher-level mineral element than dry meat. Environment factors (water resources, forage, and soil conditions) influence the qualities of Gangba lamb meat, including its mineral content (Zhang *et al.* 2021). Furthermore, Mortimer *et al.* (2014) found that genetic factors also influence lamb meat quality. There is limited data on the macro- and micromineral content of lamb meat, and there are no publications on the mineral content of Sapudi lamb meat from Indonesia.

Sapudi sheep are a genetic resource of local Indonesian livestock that are well suited to the Indonesian environment. This was based on the Minister Agriculture's Decree Number of 2389/Kpts/LB.430/8/2012 on the Determination of Sapudi Sheep Clusters in the Madura Islands and Eastern Java. The Sapudi sheep breeding center in East Java is in the Breeding Unit and HMT Livestock in Sidomulyo, Jember, and is managed by the East Java Provincial Livestock Office. Sapudi sheep have the potential to develop into meat producers with great adaptation and disease resistance. This sheep was distinguished by its large tail packed with fat and curled in shape (sigmoid). Several investigations into the peculiarities of Sapudi sheep have revealed that they possess specific qualities. According to Tanzilla (2018), Sapudi sheep are predominantly white on the head and body, males have a convex facial line (64.23%), females have a straight facial line (42.86%), almost all male cattle are hornless, 100% of female cattle are hornless, the ears are 100% upright, and the concave dorsal line is raised backward. There were three

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varieties of tail shapes, with sigmoid curves accounting for 66.67% of males and females, triangular tail shapes accounting for just 8.34% and 2.99%, and straight tail shapes accounting for 25.00% of males and 30.37% of females. Sapudi sheep had a maximum body weight of 22.09  $\pm$  4.74 kg in males and 29.56  $\pm$  3.71 kg in females. Males have a body length of 55.57  $\pm$  5.99 cm, while females have 59.62  $\pm$  3.15. Males had a chest circumference of 63.47  $\pm$  5.18 cm, while females measured 67.87  $\pm$  3.60 cm. Sapudi sheep had shoulder heights of 57.04  $\pm$  3.90 cm and 59.00  $\pm$  3.33 cm for females (Tanzilla 2018).

Previous research on the amino acid and fatty acid composition of Sapudi and Cross-merino lambs found that Cross-merino lambs had higher levels of glycine, arginine, serine, tyrosine, phenylalanine, lysine, and threonine than Sapudi lambs. A total of 51 fatty acids were discovered, with 14 and 13 types of fatty acids detected in Sapudi and Cross-Merino, respectively. Sapudi's fatty acid content is higher than that of Crossmerino's. The two lambs had a complete fat and amino acid profile. The breed determines the amino acid and fatty acid levels in lamb meat. Wang et al. (2021) found that the genetics of sheep affect quality, including nutrient content, meat compactness, and mineral content, and that this can be utilized as a criterion for selecting sheep for quality enhancement in Australia. The goal of this study was to determine the macro and micro mineral and vitamin B content of Sapudi and Cross-merino lamb meat. This study is significant because it provides fundamental information on the features of local sheep genetic resources and serves as a reference for breeding.

### **METHODS**

#### Livestock Rearing and Sampling

The sample consisted of three Sapudi sheep and three Cross-merino sheep aged one year and weighing  $25.26 \pm 1.49$  kg. Sheep were kept in uniform conditions for 2 months, with a 1-week adaptation period. The provision of complete concentrated feed is 3% of the body weight (Khasanah *et al.* 2021). Livestock slaughter was carried out halally at the Jember Slaughterhouse technical implementation unit by cutting the neck, respiratory tract, food ducts, and blood vessels of the carotid artery and jugular vein. The meat sample used for analysis was the longissimus dorsi area.

#### **Mineral Content Determination**

The mineral content was determined using inductively coupled plasma optical emission spectrometry (ICP-OES). The standard curve of the mineral mixture had six concentration points. A 0.5 g sample of meat was dipped in 600  $\mu$ L and 7.5 mL HCl in vessel tubes and allowed to stand for 15 min. The tubes were closed, and the sample was processed in a

microwave digester. The destruction results were then transferred to a 50 mL measuring flask, where a standard yttrium solution of 100 mg/L was added. The solution was then filtered through filter paper, and the intensity of the solution sample was measured using the ICP-EOS system. The wavelengths utilized were Ca: 317,933 nm; Cu: 327.395 nm; g Fe: 238.204 nm; Na: 568.821 nm; P: 214.914 nm; Zn: 213.857 nm; K: 766.491 nm; Mn: 257.610 nm; and Y: 371.029 nm (Maria *et al.* 2007). The mineral content was calculated using the following formula:

mineral content (ppm) = 
$$\frac{\frac{A_{spl}-a}{b} \times V \times fp}{W_{spl} \text{ or } V_{spl}}$$

where:

а

 $A_{\rm spl}$  = Intensity of the sample

Intercept of standard calibration curves

*b* = Slope of standard calibration curves

*fp* = Dilution factor

V = Volume of final flask sample (mL)

 $W_{Spl}$  = Sample weight (g)

 $V_{\text{Spl}}$  = Sample volume (mL)

#### Vitamin B Determination

The vitamins were B1, B2, B3, B6, and B12. Were measured using UPLC-PDA (Antakli et al. 2015), and vitamin B12 was assessed using LC-MS/MS (Ren et al. 2015). In a 10 mL amber measuring flask, vitamin B standards were prepared in series at 6-point concentrations. The sample (3 g) was placed on a 25 mL amber volumetric flask, after which NaH<sub>2</sub>PO<sub>4</sub> 0.05 M pH 6.3) was added to half of the flask capacity. The sample was sonicated for 15 min, and 25 mL of NaH<sub>2</sub>PO<sub>4</sub> (pH 6.3) was added and homogenized. The sample solution was transferred to a 2 mL tube and centrifuged at 14,000 rpm for 3 minutes. The sample was filtered via a 0.20 µm filter into a 2 mL sample vial and placed into the UPLC machine. The instrument settings included a C18 column, mobile phase A with 0.1% H<sub>3</sub>PO<sub>4</sub> and B with acetonitrile, and a gradient pump system set to 40°C. The PDA wavelengths for vitamins B1, B2, and B3 were 245 nm, 265 nm, and B6 was 283 nm, respectively. Vitamin B levels were calculated using the following formula:

$$vitamin \ B \ content = \frac{\left(\frac{Aspl-a}{b}\right) x \ FP \ x \ Va}{W_{Spl} \ or \ V_{Spl} \ x \ 10}$$

where

 $A_{Spl}$  = Sample area

*a* = Intercept of standard calibration curves

*b* = Slope of standard calibration curves

FP = Dilution factor

Va = Final volume of solution

 $W_{Spl}$  = Sample weight (g)

 $V_{\text{Spl}}$  = Sample volume (mL)

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The vitamin B12 determination was performed as cobalamin by preparing a standard series of at least 7 concentration points in a 2 mL vial and diluting them with a standard solvent. The sample (up to 1 g) was placed in a 50 mL falcon tube, followed by extracting solvent, homogenization, and sonication. The sample was centrifuged, activated using SPE, and then diluted with methanol. The eluate was evaporated using a nitrogen evaporator and then vortexed. The solution was filtered, placed in a 2 mL amber vial, and then injected into the LC-MS. The HSS T3 column of mobile phase A used 10 mM ammonium and 0.1% formic acid in distilled water and mobile phase B used 10 mM formic acid and 0.1% formic acid in methanol with a gradient pump system. Vitamin B12 levels were calculated using the following formula:

$$Cyannocobalamin \ content = \frac{(A \ spl - a)}{b} - V \ x \ FP$$
sample weight x 10

where

Aspl= Sample areaa= Intercept of standard calibration curvesb= Slope of standard calibration curvesFP= Dilution factorV= Final volume of solution

#### **Data Analysis**

The analysis data were collected and tabulated before being statistically evaluated using an independent *t*-test using SPSS 26 software (Chicago, USA).

# **RESULTS AND DISCUSSION**

### **B** Vitamins Level

B vitamins are water-soluble that is required as a coenzyme in cellular functions related to energy metabolism in the mitochondria via FMD and flavin adenine dinucleotide (FAD), which act as electron carriers (Massey 1994) and as an antioxidant in the of oxidative stress prevention (Ashoori and Saedisomeolia 2014). Table 1 displays the results of the vitamin B content analysis of Sapudi and Crossmerino lamb meat. According to the using the UPLC-PDA method, vitamins B1, B6, and B12 were not detected in either sample: nevertheless, the results of the three vitamins were below the detection limit. Vitamin B2 (riboflavin) and B3 (niacin) were found. Sapudi sheep had a much lower vitamin B2 content than Cross-merino sheep; however, there was no statistically significant difference in vitamin B3 levels. Vitamin B2 is known to work as an antioxidant, preventing oxidative stress, particularly lipid peroxidation and reperfusion oxidative injury. Furthermore, vitamin B2 has been linked to various disorders, including cancer (Yang et al. 2013), anemia (Northrop-Clewes and Thurnham 2013), CVD (Ahn et al. 2019), and cataracts (Schmidinger et al. 2013).

Niacin is typically found in two forms: nicotinic acid (NA) and nicotinamide (Gerald 2012). Niacin can also refer to various chemicals that contain nicotinic acid, nicotinamide, or molecules that have been transformed into NA. Vitamin B3 (niacin) is one of the therapeutic medicines linked with the alteration of increased HDL and decreased LDL, which are associated with high lipoprotein and hypertriglyceridemia (Mackay et al. 2012). Nicotinamide adenine dinucleotide (NAD<sup>+</sup>) is an essential vitamin B3 derivative cofactor that is phosphorylated to nicotinamide adenine dinucleotide phosphate (NADP<sup>+</sup>) and reduced to NAD(P)H. This form is a cofactor that participates in all major bioenergy activities via anabolic and catabolic pathways. The form also influences the alteration of post-translational proteins involved in cell metabolism and signaling (Makarov et al. 2019).

The levels of vitamins B2 and B3 in Sapudi and Cross-merino sheep (Indonesian sheep) were lower than those reported (Zhu et al. (2020), who found vitamins B2, B3, and B9 in the meat of Subei Gansu sheep (China sheep). Furthermore, it has been found that different portions of the muscle influence the levels of vitamins B2, B3, E, and B9 (folic acid) in meat. Vitamin B2 levels were highest in the square-cut shoulder (0.438 mg/100 g) and lowest in the breast and flap (0.177 mg/100 g). Meanwhile, the leg-chump off had the maximum vitamin B3 content (8,560 mg/100 g), whereas the topside, tripe, neck, delicate loin, short loin chop, and shoulder rack had lower levels (5,746-6,745 mg/100 g) (Zhu et al. 2020). Vitamins B1, B6, B9, and B12 were not found or were below the detection limit (B1 = 0.03 mg/100 g; B6 = 0.06 mg/100g; B9 = 50 mcg/100 g; B12 = 0.08 mcg/g).

# **Mineral Contents**

The mineral content of Sapudi and Cross-merino lamb meat differed significantly in various factors, particularly Mn and Cu levels. Mn in Cross-merino sheep was superior to that in Sapudi sheep. This is consistent with the Cu content of Cross-merino sheep, which outperformed Sapudi sheep. This study's mineral data on various observational variables, including potassium, sodium, phosphorus, selenium, zinc, iron, and calcium levels, revealed no significant differences. Table 2 shows that selenium was not detected in Sapudi sheep. It is possible that the mineral selenium is too low to be detected.

Minerals are incredibly beneficial to the human body, particularly in the formation of muscles, conditions, and enzyme cofactors. Cross-merino lamb meat includes more minerals, particularly Mn and Cu. Autukaite *et al.* (2020) found that blood mineral content varies by country/location. A detailed investigation of

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Variable	Sapudi (mg/100 g)	Cross-merino (mg/100 g)
Vitamin B2 (Riboflavin)	0.11 ± 0.01 <sup>a</sup>	$0.20 \pm 0.01^{b}$
Vitamin B3 (Niacin)	0.51 ± 0.01 ª	$0.40 \pm 0.13^{b}$

Remark: Different superscripts showed significant differences at the 1% level (p<0.01).

Table 2 Mineral content of Sapudi and Cross-merino lamb

Variable	Sapudi (ppm)	Cross-merino (ppm)
Potassium	250.10 ± 14.471	258.37 ± 1.31
Sodium	69.53 ± 14.79	67.12 ± 9.52
Phosphorus	1431.46 ± 73.53	1468.89 ± 24.45
Calcium	91.40 ± 5.59	99.86 ± 8.51
Vanganese	$0.45 \pm 0.17^{a}$	1.44 ± 0.77 <sup>b</sup>
Selenium	Not detected*	12.78 ± 0.96
Zink	$4.10 \pm 0.70$	$4.22 \pm 0.58$
Iron	1.96 ± 0.11	2.21 ± 0.51
Copper	$4.25 \pm 0.06^{a}$	$5.04 \pm 0.50^{b}$

Remark: Different superscripts show significant differences at the 1% level (p<0.01).

the iron and copper content of Australian lamb (part of *longissimus lumbrum*) found that the amounts of both minerals were 2.03 mg/100 g and 2.43 mg/100 g (Mortimer *et al.* 2014). Further research in Australian sheep discovered the mineral copper at 3.5 mg/100 g.

Breeding attempts to boost lean meat yield have resulted in a decrease in Fe content and an increase in Mg content (Knight et al. 2020). In addition to genetic considerations, the minerals in meat are controlled by the nutrition consumed by sheep (Mortimer et al. 2014). The mineral composition of meat is also influenced by the slaughter site and age of the sheep. According to Lin et al. (1988), the mineral concentration increases with increasing age. They also found that the foreshank and shoulder regions contained more zinc than the other components. Furthermore, Osorio et al. (2007) revealed that cattle fed milk replacers and suckling lambs performed differently in terms of mineral content in meat, with cattle breastfed by their mothers containing more potassium, phosphorus, and copper, and less zinc and manganese (Table 2). Our findings show that the nation has a considerable impact on the mineral and vitamin content of lamb meat. However, the composition of carcasses varies substantially depending on numerous aspects, including rearing system, nutrition, ethnicity, age, sex, slaughter weight, and pre-slaughter management (Corazzin et al. 2019).

### CONCLUSION

Sapudi sheep meat contains less vitamin B2 but more vitamin B3, less manganese and copper than Cross-merino. The vitamin and mineral composition of lamb meat varies depending on the breed. Local sheep breeds, which are genetic resources of Indonesia, can be used to provide nutritious and high-quality sheep meat.

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# REFERENCES

- Ahn JY, Kim IS, Lee JS. 2019. Relationship of riboflavin and niacin with cardiovascular disease. *Korean Journal of Clinical Laboratory Science*. 51(4): 484– 94. https://doi.org/10.15324/kjcls.2019.51.4.484
- Antakli S, Sarkees N, Sarraf T. 2015. Determination of water-soluble vitamins B1, B2, B3, B6, B9, B12 and c on C18 column with particle size 3 µM in some manufactured food products by hplc with UV-DAD/FLD detection. *International journal of pharmacy and pharmaceutical Sciences*. 7(6): 219– 224.
- Ashoori, Marziyeh, Saedisomeolia A. 2014. Riboflavin (vitamin B2) and oxidative stress: a review. *British Journal of Nutrition*. 111(11): 1985–1991. https://doi.org/10.1017/S0007114514000178
- Autukaitė J, Poškienė I, Juozaitienė V, Undzėnaitė R, Antanaitis R, Žilinskas H. 2020. Influence of season and breed on serum mineral levels in sheep. *Polish Journal of Veterinary Sciences.* 23(2): 473–476. https://doi.org/10.24425/pjvs.2020.134695
- Belhaj K, Farid M, Moumen AB, Sindic M, Fauconnier ML, Boukharta M, Caid HS, Elamrani A. 2021. Proximate composition, amino acid profile, and mineral content of four sheep meats reared extensively in Morocco: a comparative study. *Scientific World Journal.* 2021. https://doi.org/10.1155/2021/6633774

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- Corazzin M, del Bianco S, Bovolenta S, Piasentier E. 2019. Carcass Characteristics and Meat Quality of Sheep dan Goat. More than Beef, Pork and Chicken
  The Production, Processing, and Quality Traits of Other Sources of Meat for Human Diet. 119–165. Switzerland (CH): Springer, Cham. https://doi.org/10.1007/978-3-030-05484-7\_6
- Ekmekcioglu C, Wallner P, Kundi M, Weisz U, Haas W, Hutter HP. 2017. Red meat, diseases, and healthy alternatives: a critical review. *Critical review in food Science and Nutrition.* 58(2): 247–261.
- Jacobson MK, Jacobson EL. 2018. Vitamin B3 in health and disease: toward the second century of discovery. *Methods in Molecular Biology*. 1813: 3– 8. https://doi.org/10.1007/978-1-4939-8588-3\_1
- Khasanah H, Syaikhullah S, Adhyatma M. 2022. The carcass and nutritional meat characteristics of sapudi and cross-merino sheep. *Buletin Peternakan.* 46(3): 184–192. https://doi.org/10.21059/buletinpeternak.v46i3.740 48
- Knight MI, Butler KL, Linden NP, Burnett VP, Ball AJ, McDonagh MB, Behrendt B. 2020. Understanding the impact of sire lean meat yield breeding value on carcass composition, meat quality, nutrient and mineral content of australian lamb. *Meat Science*. 170 (December): 108236. https://doi.org/10.1016/j.meatsci.2020.108236
- Lin KC, Cross HR, Johnson HK, B. C. Breidenstein, V. Randecker, RA. Field. 1988. Mineral Composition of Lamb Carcasses from the United States and New Zealand. *Meat Science* 24(1): 47–59.
- Mackay D, Hathcock J, Guarneri E. 2012. Niacin: chemical forms, bioavailability, and health effects. *Nutrition Reviews*. 70(6): 357–66. https://doi.org/10.1111/j.1753-4887.2012.00479.x
- Makarov MV, Trammell SAJ, Migaud ME. 2019. The chemistry of vitamin B3 metabolome. *Biochemical Society Transactions*. 47(1): 131–47. https://doi.org/10.1042/BST20180420
- Mioč B, Vnučec I, Prpić Z, Pavić V, Antunović Z, Barać Z. 2009. Effect of breed on mineral composition of meat from light lambs. *Italian Journal of Animal Science. 8*(sup3): 273–275.
- Mlambo V, Mapiye C. 2015. Towards household food dan nutrition security in semi-arid areas: what role for condensed tannin-rich ruminant feedstuffs. *Food Research International.* 76(P4): 953–961.
- Mortimer SI, van der Werf JHJ, Jacob RH, Hopkins DL, Pannier L, Pearce KL, Gardner GE, Warner RD, Geesink GH, Edwards JEH, Ponnampalam EN, Ball AJ, Gilmour AR, Pethick DW. 2014. Genetic parameters for meat quality traits of australian lamb

meat. *Meat Science.* 96(2): 1016–1024. https://doi.org/10.1016/j.meatsci.2013.09.007

- Northrop-Clewes CA, Thurnham DI. 2013. Biomarkers for the differentiation of anemia and their clinical usefulness. *Journal of Blood Medicine*. 20(4): 11– 12. https://doi.org/10.2147/JBM.S29212
- Osorio MT, Zumalacárregui JM, Bermejo B, Lozano A, Figueira AC, Mateo J. 2007. Effect of ewe's milk versus milk-replacer rearing on mineral composition of suckling lamb meat and liver. *Small Ruminant Research*. 68(3): 296–302. https://doi.org/10.1016/j.smallrumres.2005.11.010
- Rekanović S, Grujić R, Vučić G, Hodžić E. 2019. Mineral composition of traditional sheep meat products depends on the thermal treatment. *Journal* of Hygienic Engineering and Design. 29: 92–98
- Ren XN, Yin SA, Yang ZY, Yang XG, Shao B, Ren YP, Zhang J. 2015. Application of UPLC-MS/MS method for analyzing b-vitamins in human milk. *Biomedical and Environmental Sciences: BES*. 28(10): 738–750.
- Schmidinger G, Pachala M, Prager F. 2013. Pachymetry Changes during Corneal Crosslinking: Effect of Closed Eyelids and Hypotonic Riboflavin Solution *Journal of Cataract & Refractive Surgery*. 39(8): 1179–1183.
- Gerald F C Jr. 2012. *The Vitamins (Chapter 12-Niasin)*. Paris (FR): Academic Press.
- Tanzilla M. 2018. Karakteristik sifat kualitatif dan kuantitatif domba sapudi h. [Undergraduate thesis]. Malang (ID): Universitas Brawijaya.
- Wang Q, Liu H, Zhao S, Qie M, Bai Y, Zhang J, Guo J, Zhao Y. 2021. Discrimination of mutton from different sources (regions, feeding patterns and species) by mineral elements in inner mongolia, china. *Meat Science*. 174(April): 108415. https://doi.org/10.1016/j.meatsci.2020.108415
- Yang HT, Chao PC, Yin MC. 2013. Riboflavin at high doses enhances lung cancer cell proliferation, invasion, and migration. *Journal of Food Science*. 78(2): H343–H349.
- Zhang Q, Que M, Li W, Gao S, Tan X, Bu D. 2021. Gangba sheep in the tibetan plateau: validating their unique meat quality and grazing factor analysis. *Journal of Environmental Sciences.* 101(March): 117–122.
- Zhu H, Qiu J, Liang KH, Wang J. 2020. Variability in nutritional profile and textural properties of different cuts of lamb meat from Subei Gansu. In: Proceeding of an International Conference on Agricultural Science and Technology and Food Engineering (ASTFE 2020). Online. Changchun, September 15<sup>th</sup>, 2020.

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